## An Overview of Agarwood, Phytochemical Constituents, Pharmacological Activities, and Analyses

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## ABSTRACT

Agarwood is a resin-impregnated heartwood obtained from the plants belongs to the genera, Aquilaria, Daphne, Gonystylus, Gyrinops and Wikstroemia. It is traditionally used for the production of perfume and incense stick, and pharmaceutical applications. Agarwood usually induced by the natural (traditional), conventional, and non-conventional methods. The major groups of phytochemicals identified in agarwood extracts are sesquiterpenes, 2-(2-phenylethyl)-4H-chromen-4-one derivatives (PECs), and aromatic compounds. These phytochemicals are showed various pharmalogical properties such as anti-inflammatory, cytotoxic, neuroprotective, antidiabetic, anti-bacterial, etc. Several analytical techniques are applied to analyze the agarwood phytochemicals including sesquiterpenes, which exists mostly in the form of essential oils, and the fragrance constituents of PECs. The present review summarize the agarwood traditional uses, induction methods, phytochemical constituents, potential pharmacological activities, along with analyses methods. This review was carried out by searching various scientific databases, including Google Scholar, PubMed, Elsevier, ACS publications, Taylor and Francis, Wiley Online Library, MDPI, Springer, Thieme, and ProQuest. The present review provides a scientific basis for future studies and necessary information for the development of agarwood based therapeutic agents.

**Keywords:** Agarwood; Traditional uses; Induction methods; Chemical constituents; Biological activities; Analyses

## INTRODUCTION

Agarwood, known as aloeswood or eaglewood, is an aromatic dark resin-impregnated heartwood obtained from wounded tree species of the Thymelaeaceae family [1]. The plant family Thymelaeaceae contains 54 genera, including *Aquilaria*, *Daphne, Gonystylus, Gyrinops and Wikstroemia* [1]. The species of the genus *Aquilaria, Gonystylus*, and *Gyrinops* produce agarwood [1]. In particular, the genus *Aquilaria* contains 57 species, among these 21 are accepted in the plant list [1]. So

far, fifteen species of *Aquilaria* and nine species of Gyrinops are reported as agarwood producing plants [2]. Agarwood (resin)producing species are found from the forests of Southeast Asia including, Bangladesh, Bhutan, China, India, Indonesia, Laos, Malaysia, Myanmar, Singapore, Taiwan, Thailand, and Vietnam [1,2]. They are usually found in lowland tropical forests with optimal sunlight, shade and moisture. Agarwood-producing species have a small flower similar to that of 'jasmine', and the fruit is bitter [3].

Healthy Aquilaria tree does not produce agarwood [2]. The healthy wood is white, soft, even-grained and not having a perfumed smell, as compared with the dark, hard and heavy scented characterictics resin-impregnated agarwood [2]. The agarwood resin developed through pathological, wounding and non-pathological mechanisms [4]. The formation of agarwood occurs naturally in response to natural injuries such as lightning, insects and mold attacks [4]. The deposited resin around the wounds over the years accumulate and eventually forms agarwood [4]. Therefore, Agarwood is termed as the resin-impregnated pieces of wood [4], and its formation is related to the self-defense mechanism of Aquilaria trees in response to biotic and abiotic stresses [1,2]. Stresses trigger the defense responses of Aquilaria species, which in turn initiate the secondary metabolite biosynthesis and the accumulation of agarwood resin [1,2].

The prominent species of agarwood producing Aquilaria species are, A. beccariana Tiegh., A. crassna Pierre ex Lecomte, A. filaria (Oken) Merr., A. hirta Ridl., A. khasiana Hallier f., A. malaccensis Lamk., A. microcarpa Baill., A. rostrata Ridl., A. sinensis (Lour.) Spreng., and A. agallocha [5]. Among these A. agallocha, A. crassna, A. malaccensis, and A. sinensis gain significant attention due to their therapeutic uses in traditional Southeast Asian medical systems [5]. Accordingly, these species appear frequently in the literature, particularly A. crassna, A. malaccensis and A. sinensis [1,5]. The agarwood producing Aquilaria species and their native place are presented in Table 1. With the increasing demand for agarwood, the population of agarwood species is declining rapidly in the wild. Currently, the genus Aquilaria is listed as endangered species and protected under Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES) regulation [1]. The index of CITES species listed A. agallocha Roxb. as a synonym of A. malaccensis Lamk [1]. A. malaccensis Lamk., is also synonym to Aquilariella malaccensis (Lam.) Tiegh., and Agallochum malaccense (Lam.) Kuntze [6]. The International Union for Conservation of Nature and Natural Resources (IUCN) Red List of Threatened Species is listed A. crassna as critically endangered, and A. malaccensis and A. sinensis as vulnerable [7].

Species	Place of origin (Native)
Aquilaria apiculata Merr	Philippines
<i>Aquilaria baillonii</i> Pierre ex Lecomte	Cambodia, Thailand, Laos, Vietnam
Aquilaria banaensis P.H.H6	Vietnam
Aquilaria beccariana Tiegh	Indonesia
<i>Aquilaria citrinicarpa</i> (Elmer) Hallier f.	Philippines
<i>Aquilaria crassna</i> Pierre ex Lecomte	Thailand, Cambodia, Vietnam
<i>Aquilaria cumingiana</i> (Decne) Ridl	Malaysia
Aquilaria khasiana Hallier f.	India
Aquilaria malaccensis Lam.	India, Myanmar, Malaysia, Indonesia, Philip- pines
Aquilaria microcarpa Baill	Indonesia
<i>Aquilaria parvifolia</i> (Quisumb) Ding Hon	Philippines
Aquilaria rostrata Ridl	Malaysia
Aquilaria rugosa Kiet Kessler	Vietnam

Aquilaria sinensis (Lour.) Gilg	China
Aquilaria subintegra Ding Hon	Thailand
Aquilaria urdanetensis (Elmer) Hallier f.	Philippines
Aquilarla yunnanensis S.C. Huang	China

Table 1: Agarwood producing Aquilaria species and their place of origin.

#### 1.1. Traditional uses

Agarwood is known as "wood of God" because of religious practices [4]. The word "aloes" which means agarwood is found in the Sanskrit poet, Kâlidâsa, dated back to c. 4th-5th century CE [4]. Agarwood is considered the finest natural incense and has been used in many cultures, such as the Arabian, Chinese, Indian, and Japanese cultures [8]. Agarwood also associated with religious history, rituals and ceremonies in Buddhism, Christianity, Hinduism, and Islam [8]. It is known as gaharu in the Indonesia and Malaysia, jin-koh in Japan, chen xiang (沉香) in Chinese, agar in India, chim-hyuang in Korea, kritsana noi in Thailand, tram huong in Vietnam, and oud in the Middle East [8]. Agarwood is widely used as therapeutic perfumes, traditional medicine, religious purposes and aromatic food ingredient (Table 3) [3]. In the traditional Chinese and Ayurvedic medicines as an aphrodisiac, sedative, cardiotonic and carminative, as well as to treat gastric problems, coughs, rheumatism and high fever [9]. Agarwood is a traditional Chinese medicine included in the 2020 edition of Chinese Pharmacopoeia [10]. In traditional Arabian medicine, agarwood essential oil is used for aromatherapy [3]. In Thailand, agarwood has been used for a long time as a traditional treatment for infectious diseases such as diarrhea and skin diseases [3]. Additionally, *A. crassna* extract has been using as the ingredient of Ya-hom, a traditional Thai herbal formulation for the treatment of fainting by targeting the cardiovascular system.

#### 1.2. Grading system

The market price of agarwood has commercial attention. However, the grading process of agarwood is largely depends on the human experience from the age-old practices of each country [1]. In general, the classification of agarwood oil quality is based on wood physical properties, long lasting aroma when burnt, color, resin content, high fixative properties and consumer perception, etc [1]. The higher the grade of agarwood, the richer the layers of aroma [1]. The best agarwood fragrance is mellow and sweet, full of penetration and persistence, and the powdery waxy material on the surface can be scraped off and kneaded it into a ball [1]. Its aroma is regarded as a symbol of high quality. Complexity and variability in agarwood composition are major challenges associated with its grading process. The morphological grading system of agarwood is shown in Table 2.

Observational Feature	Grading Categary			
Observational realure	A	В	С	
Sense of oiliness	Strong	Strong	Mild	
Aroma	Strong, feel sweet and cool	Less potent odor, feel	Mild aroma, feel slightly	
	5, 11, 11, 11, 11, 11, 11, 11, 11, 11, 1	sweet and slightly spicy	sweet, salty	
Resin density	High dense, compact, sink	Dense, less compact,	Light and not dense,	
Resilidensity	in water when soaked	half-sinkage in water	full-floating on water	
Weight	Hard texture, brittle, and	Texture little hard, little	Loose texture, not brittle,	
weight	not hollowed	brittle, slightly hollow	and hollow	

Table 2: Assesment of agarwood quality using grading system.

#### 1.3. Economical value

Agarwood is a valuable, non-timber forest product used different societies for medicinal, aromatic, cultural and religious purposes [8]. As the wealth of the consumer countries are gradually increased in recent decades, the market's demand for agarwood started to exceed its supply [1]. The market value of agarwood derivative products is dependent on the classification or grading of agarwood, which is determined by a cumulative factor of the fragrance strength and longevity, resin content, geographical origin and purity (for oil) [8]. Global agarwood prices can be ranging from US\$20 - 6,000 per/kg for the wood chips depending on its quality or US\$ 10,000 per/kg for the wood itself [1]. High quality wood is used as incense in Arabian households and for the 'koh-doh' incense ceremony in Japan [8]. High-quality agarwood products can reach prices as high as US\$100,000/kg. In the form of oud oil which is distilled from agarwood for perfumery, can be sold for US \$1500 per 11.7 g [1,9]. The annual global market for agarwood has been estimated to be in the range of US\$ 6 – 8 billion [1,9]. Agarwood has commercial importance in three categories i.e. perfume production, incense stick, and pharmaceuticals as described below [8].

#### 1.3.1. Perfume

Agarwood oil is an essential oil obtained by water and steam distillation of agarwood [1]. Agarwood oil is a yellow to dark amber, viscous liquid with a characteristic balsamic and woody odour [9,11]. It is used in luxury perfumery for application. Agarwood perfumes are commonly prepared in both alcoholic and non-alcoholic carriers [9,11]. Agarwood perfume has a unique smell obtained from fragrance essential oil and aromatic compound [11]. The oil is also used as a fragrance in the production of cosmetics and personal care products, such as soaps and shampoos [9,11]. Agarwood resin is a key ingredient in old and new Arabic perfume products, and used as an element within high-quality perfumes in Arabic, Japanese and Indian cultures [4]. Traditionally in the Middle East, agarwood oil is used as a scent, and Minyak attar (water-based) [4]. "Attar" is an example of a water-based perfume containing agarwood oil, which is traditionally used by Muslims to lace prayer clothes [4]. Agarwood oil is one of the most important ingredients in Chinese perfume industry; additionally it became a prominent in the modern western perfume and fragrance industry.

#### 1.3.2. Incense stick

Burning agarwood produces fragrance, which is used as incense for ceremonial purposes in Buddhism, Confucianism and Hinduism [11]. The incense also functions as an insect repellent [4,8]. The aromatic compounds are the main chemical components in agarwood smoke and create an atmosphere of peace and serenity [1,4]. It scent heavenly, woody nuance, balsamic and warm aura of bittersweet when the chromones break into low molecular weight at high temperature [1,4]. In Taiwan, the agarwood stick is used in traditional festivals or ceremonies to bring safety and good luck to the believer [11]. The agarwood incense stick is used in the bathroom as a customary sense, during Ramdan prayer by the Muslim, and Puja celebration by the Hindu religious practice [11].

#### 1.3.3. Pharmaceutical use

Agarwood plays a vital role in the field of medicine, contains various chemical components, including several sesquiterpenes, 2-(2-phenylethyl)chromens (PECs), and aromatic compounds, etc [3,6,12]. These compounds display various biological properties such as anticancer, anti-inflammatory, antioxidant, antibacterial, antifungal, antidiabetic, and so on [3,6,12]. Traditionally agarwood is prescribed to treat pleurisy by the Sahih Muslim, relieve pain, arrest vomiting, and asthma [6,12]. A. malaccensis products are an essential source in the field of Ayurveda for treating various diseases such as appetizer, analgesic, antipyretic, antihistaminic, styptic, carminative, cytotoxic, insecticidal, general tonic, etc [6,11,12]. Agarwood materials have also been formulated into a balm (muscle rub) and candle wax [11]. The pharmaceutical and traditional use of agarwood in different countries/locations are presented as in Table 3.

Place	Traditional use	Preparations/route of intake
Bangladesh	Treatment of rheumatism	Agarwood taken orally
China	Treatment of circulatory disorders, abdominal pain, vomiting, dyspnea, asthma	Heartwood in Chinese medicines, and Heartwood decoction
India	Treatment of diarrhoea, dysentery, vomiting, anorexia, mouth and teeth diseases, facial paralysis, shivering, sprains, bone fracture	Heartwood in Ayurvedic formulation such as Chawanprash, Arimedadi Taila and Mahanarin Taila
Indonesia	Treatment of joint pain, Sedation, detoxification, treat- ment of stomachaches, incense sticks	Wood burned and smoke held over the affected area
Japan	Stomachic and sedative agent	Infusion or decotion
Korea	Treatment of cough, acroparalysis, croup, asthma, sto- machic agent, tonic, sedative and expectorant	Infusion or decotion
	Tonic, stimulant and carminative agent after childbirth	Heartwood mixed with coconut oil
Malaysia	Treatment of rheumatism and body pains	Heartwood decoction (mixed with other types of woods)
	Treatment of small pox	Heartwood prepared into Ointment
	Stop bleeding of the wounds	Bark and roots
Philippines	Treatment of malaria (substitute for quinine)	Bark, wood and fruits
Thailand	Treatment for diarrhoea, dysentery and skin diseases, antispasmodic and cardiovascular function enhancer in fainted patient	Traditional medicinal preparation 'Kris- anaglun
	Treatment of fainting, nausea and vomiting	Folk medicine 'Ya–Hom'
Tibet	Treatment of nervous and emotional disorders Cardioprotective agents	Infusion or decotion

**Table 3:** The pharmaceutical and traditional use of agarwood in different countries/locations.

#### 1.3.4. Other uses

The uses of agarwood is not restricted to incense and perfumery. Solid pieces of agarwood are carved into natural art sculptures, beads, bracelets and boxes [4,11]. The wood of *A. agallocha* is used as decorative ornaments (China), 'joss sticks' (China and India), and flea and louse repellents (India), whereas the bark has been used to manufacture paper (China) [1,4]. In India, the wood of *A. malaccensis* used as fuel for fumigation, and the bark has been used to make cloth and rope [11].

#### 1.4. Agarwood induction methods

Agarwood is a valuable non-timber product, and its demand is much greater than its supply. The agarwood (resin) induction mechanism is not fully understood or elucidated. High demand of quality agarwood in conjunction with the depletion of the wild *Aquilaria* trees, leads to the artificial induction of agarwood resin formation. Modern artificial agarwood formation techniques are mainly biochemical methods, such as chemical reagent invasion and bacteria inoculation (Figure 1) [2].

#### 1.4.1. Natural (Traditional) methods

Naturally, agarwood formation is often linked to the physical wounding or damage of *Aquilaria* trees caused by thunder strike, animal grazing, pest and disease infestations [13]. These events expose the inner part of the trees toward pathogenic microbes, which evoke the defense mechanism of *Aquilaria* to initiate the resin production [13]. This natural formation process of agarwood has greatly inspired the development of diverse artificial induction methods (Figure 1). For example, the microbial species of *Actinobacteria* sp., *Acidobacteria* sp., *Aspergillus* sp., *Alcaligenes* sp., *Bacillus* sp., *Chaetomium* sp., *Curvularia* sp. *Fusarium* sp. *Lasiodiploidia* sp., *Penicillium* sp., *Proteobacteria* sp., *Pseudomonas* sp., and *Trichoderma* sp. are involved in the agarwood formation [1,13]. For more details, please refer the recent review article [221].

#### 1.4.2. Conventional methods

Various conventional methods are applied to initiate agarwood resin formation. The techniques often involved the physical penetration into the trunk (wounding), mechanical wounding, axe chopping, nailing, holing, burning, insertion of a microbial (mainly fungal) concoction (pathology) and response of the tree towards the administered stress (non-pathological) [2,5]. Many pure-culture strains of fungi such as *Aspergillus niger*, A. fijiensis, Chaetomium sp., Fusarium solani, Lasiodiplodia sp. (L. hormozganensis), Gongronella butleri, Saitozyma podzolica, Cladorrhinum bulbillosum, Humicola grisea, Penicillium sp., Trichoderma lentiforme, Phaeoacremonium rubrigenum, and Tetracladium marchalianum are isolated from natural agarwood are found to be effective biological agents to induce agarwood formation in healthy Aquilaria trees [14-16]. For more details, please refer the recent review article [221]. Therefore, fungal-interaction induction methods coupled with the application of biological inoculum are developed for agarwood induction [2,5]. The advantage of using fungal inoculum is that it is generally believed to be safe for handling and eco-friendly. However, fungal inoculation will normally give rise to localized and inconsistent quality of agarwood due to the different fungal consortium used [2,5]. As a solution, laborious holing process and long incubation time is required to maximize the colonized surface area on the tree to produce better quality of agarwood [17]. The fungal infected Aquilaria trees are reported to deposit agarwood resin around the infected sites as barrier to prevent further fungal intrusion [13,14]. Agarwood resin deposition accompanied with color changes of internal tissues occured within a year by injuring the trees [8]. Although it is cost effective and requires only personnel with little or no scientific knowledge on agarwood, but these conventional induction methods usually result in inferior quality and uncertain yield of agarwood. Mass cultivation and large plantation of Aquilaria trees using these conventional methods have greatly resolved the shortage of agarwood supply in the global market.

#### 1.4.3. Non-conventional methods

Artificial induction of agarwood formation is the use of chemical, insect and pathogen-inducing techniques is increasingly common in agarwood induction [18]. Chemical inducers normally comprise of phytohormones, salts, minerals and biological-derived substances [2,18,19]. Various chemical induction approaches are developed, including cultivated agarwood kit (CA-kit), the whole-tree agarwood inducing technique (Agar-Wit) and biologically agarwood-inducing technique (Agar-bit). CA-kit is a combined method based on physical wounding and chemical induction, where the inducing agent is applied into the Aquilaria tree via an aeration device inserted into the wound [2]. Agar-Wit is a transpirationassisted chemical treatment to form an overall wound in the tree, where the preloaded inducer in a transfusion set is distributed via plant transpiration [18]. Similarly, Agar-bit method adopts the idea of distributing the inducing reagent

by plant transpiration, except that the reagents are injected directly into the stems of the tree [20]. Chemical inducers are suitable for mass production of agarwood with easier quality control than biological inoculum. However, in spite of the fast results and high yields, the application of chemical inducers still poses skepticism of toxicity on both human and environment. All of these induction techniques in any case mimic the natural processes of agarwood formation, which have their own strengths and weaknesses. The agarwood induction methods are presented in Figure 1. On the other hand, *in vitro* culture of various parts of *Aquilaria* spp. and *Gyrinops* spp. are studied at various tissue culture laboratories [2]. The tissue culture techniques identified the key regulator genes of *Aquilaria* spp. and *Gyrinops* spp. involved in the agarwood production [2].



Figure 1: Schematic presentation of Agarwood induction techniques.

#### 2. PHYTOCHEMICAL CONSTITUENTS OF AGARWOOD

The chemical constituents of healthy Aquilaria trees without resin-formation differ from the resin-impregnated portions of the plants [6,12]. The phytochemical analysis of agarwood resin has been the subject of many studies [1,6,12]. The types and derivatives of chemical constituents in agarwood are extremely wide and diverse, indicating the different types of fragrance properties of agarwood from different species and regional sources [1,6,12]. Agarwood resin constituents were isolated using solvent extraction, with subsequent purification via column chromatography and structural elucidation using spectroscopic techniques, including NMR [1,6,12]. Essential oils are produced by the hydrodistillation of resin followed by GC-MS or the newer technique of supercritical fluid extraction (SFE) [21]. The chemical constituents in agarwood may vary considerably in terms of quality, source plant origin, extraction methods, agarwood induction method, or agarwoodformation process, collection time, analytical approach etc [1,6,12]. The agarwood chemical constituents produced by Aquilaria species including A. sinensis, A. malaccensis (syn. A. agallocha), A. crassna, A. filaria, and Gyrinops salicifolia, as well as an unidentified Aquilaria spp [1,6,12]. Previous chemical investigations of agarwood species resulted in the isolation and structure characterization of several sesquiterpenes, 2-(2-phenylethyl)-4H-chromen-4-one derivatives (PECs), and aromatic compounds are the main characteristic chemical constituents [6,12,22]. The types of agarwood chemical constituents are described below.

#### 2.1. Sesquiterpenoids

Sesquiterpenes are composed of three isoprene units. They are mainly distributed in plants existing mostly in the form of volatile constituents present in essential oils. The constituents of agarwood essential oil is mainly composed of sesquiterpenoids, and low abundant of volatile aromatic metabolites, which gives an unique and fragrant-smelling property of agarwood [1]. The sesquiterpenes isolated from agarwood exhibit various types (Figure 2), including acoranes (A), agarospiranes (B), cadinanes (C), eudesmanes (D), eremophilanes (E), guaianes (F), humulanes (G) and prezizaanes (H), zizaanes (I).



Figure 2: Types of sesquiterpenes from agarwood.

#### 2.1.1. A. Acoranes

the agarwood of *A. sinensis* (Figure 3). The compounds **A2** and **A3** are a pair of stereoisomers.

#### The spiro sesquiterpenes, acoranes (A1-A3), are reported from



Figure 3: Chemical structures of acorane-type sesquiterpenes from agarwood.

#### 2.1.2. B. Agarospiranes (vetispiranes)

The spirocyclic sesquiterpenes, agarospiranes are reported in agarwood from *A. sinensis*, *A. malaccensis* and *A. agallocha* (Figure 4, and Table 4). The first agarospirane sesquiterpene discovered in agarwood is agarospirol (**B1**) from the agarwood of *A. agallocha* [23]. The allyl ether 2,14-epoxy-vetispir-6ene (B10) and enol ether 2,14-epoxy-vetispira-6(14),7-diene (B11) are reported from the essential oil of *A. agallocha* [24]. Vetispira-2(11),6(14)-dien-7-ol (**B8**) and vetispira-2(11),6dien-14-al (**B9**) might be artefacts [25]. The sesquiterpenes, agarospiranes have limited distribution and are mainly found in the agarwood species of *A. agallocha*, *A. malaccensis*, and *A. sinensis* (Figure 4, Table 4). Phytochemical examination of 95% ethanol extract of *A. agallocha* agarwood, resulted in the isolation of agarospirane-type sesquiterpenes (agarospiranic aldehyde A, and B, **B13**, **B14**) [26].



Figure 4: Chemical structures of agarospirane-type sesquiterpenes from agarwood.

No.	Name	Source	Ref.
		A seelleshe	27.22
<b>D1</b>	A	A. agallocha	27,23
B1	Agarospirol	A. malaccensis	28
		A. sinensis	29
B2	Baimuxinol	A. sinensis	30
B3	Baimuxinic acid	A. sinensis	30
		A. sinensis	29, 31,32
B4	Baimuxinal [Oxoagarospirol]	A. malaccensis	
D4	Baimuxinai [Oxoagarospiroi]		33,34, 35
		A. agallocha	
B5	(4 <i>R</i> ,5 <i>R</i> ,7 <i>R</i> )-1(10)-spirovetiven-11-ol-2-one	Kyara-Vietnam	36
B6	2-Oxo-12-hydroxy-hinesol	A. sinensis	37
B7	Isoagarospirol		25
B8	Vetispira-2(11),6(14)-dien-7-ol	A. agallocha	24
B9	Vetispira-2(11),6-dien-14-al	A. agallocha	24
B10	2,14-Epoxy-vetispir-6-ene	A. agallocha	24
B11	2,14-Epoxy-vetispira-6(14),7-diene	A. agallocha	24
B12	rel-(2R,5R,10S)-6(7)-Spirovetiven-11,12,13-triol	Aquilaria spp.	38

Table 4: Agarospirane-type sesquiterpenes from agarwood.

#### 2.1.3. C. Cadinanes

Two (**C1** and **C2**), decalin skeleton containing cadinanetype bicyclic sesquiterpenes are reported from agarwood of *A. sinensis* (**C1**) [39], and *A. crassna* (**C2**) [40], respectively. These two sesquiterpenes differ from eudesmane-type sesquiterpenes by the position of the isopropyl substituents and two methyl groups (Figure 5).



**Figure 5:** Chemical structures of cadinane-type sesquiterpenes from agarwood.

#### 2.1.4. D. Eudesmanes (selinanes)

The main types of sesquiterpene found in agarwood are eudesmane-type sesquiterpenes, which are a class of bicyclic sesquiterpenes with a decalin skeleton. These compounds are widely distributed in the agarwood species of *A. agallocha, A. crassna, A. malaccensis*, and *A. sinensis*, as well as in *G. salicifolia* [12,25,416]. The eudesmane-type sesquiterpenes of agarwood are presented as Figure 6, and Table 5. Most of agarwood

eudesmanes (D1-D36) contains an isopropenyl group or 2-hydroxyisopropyl group at the C-7 position, while the methyl groups at C-4 or C-11 are often oxidized to form CHO, COOH, or CH<sub>2</sub>OH groups. The eudesmanes (D3, D6, D7, D11, E19, E20 and E27) possessing an oxidation at C-9 or C-15, and an isopropenyl group at the C-7 position are reported from the acetone extract of the Vietnamese agarwood called kanankoh (A. agallocha) [35,42]. The sesquiterpenes, agarofurans, valencanes and agarospiranes (vetispiranes) biosynthetic precursor (-)-10-epi-y-eudesmol (D21) is isolated from A. malaccensis [33]. The nor-eudesmane derivatives D37-D40 are reported from the commercial agarwood oil (A. agallocha) [43]. The agarofuran sesquiterpenes D41–D55 has a trans-decalin structure, and a  $\beta$ -oriented isopropoxy bridge [12,44]. The compounds D44, D48, D49, D51, D53 and D54 are isolated from the agarwood of A. agallocha [45,46]. The sesquiterpenes D41, D43, D45, and D46 are obtained from the volatile oil of A. sinensis [47–49]. The nor-agarofuran derivatives (D52, D54 and D55, which lack the methyl group at C-4 are only reported from agarwood of A. agallocha [43,46]. A recent study reported that the phytochemical examination of 95% ethanol extract of A. agallocha agarwood, resulted in the isolation of eudesmane-type sesquiterpenes (agalleudesmanol A-I, D56-D64) [26]. Chemical examination of the ethyl ether extract of Aquilaria spp. collected in Thailand, resulted in the isolation and structure determination of eudesmane sesquiterpenes, D65, D66, and D67 [50].

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Figure 6: Chemical structures of eudesmane-type sesquiterpenes from agarwood.

No.	Name	Source	Ref.
D1.	Agarol- [11(13)-Eudesmen-12-ol]	A. agallocha	51
D2.	15-Hydroxyl-12-oxo-α-selinen	A. sinensis	52
D3.	Selina-3,11-dien-14-ol	A. agallocha	23
	12,15-Dioxo-α-selinen	A. sinensis	32,52,53
D4	[Selina-3,11-diene-12,15-dial]	G. salicifolia	54
D5	(4aβ,7β,8aβ)-3,4,4a,5,6,7,8,8a-Octahydro-7-[1-(hydroxymethyl)	A. malaccensis	55
5	ethenyl]-4a-methylnaphthalene-1-carboxaldehyde	A. sinensis	30,31,52
D6	Selina-3,11-dien-14-oic acid	A. agallocha	42
D7	(–)Selina-3,11-dien-14-al	A. agallocha	42
D8	(5S,7S,9S,10S)-(+)-9-hydroxy-selina-3,11-dien-12-al	A. sinensis	31,52
D9	(5S,7S,9S,10S)-(+)-9-hydroxy-eudesma-3,11(13)-dien-12-methyl es- ter	A. sinensis	31,52
D10	(5S,7S,9S,10S)-(–)-9-hydroxy-selina-3,11-dien-14-al	A. sinensis	52
D11	(5S,7S,9S,10S)-(+)-selina-3,11-dien-9-ol	A. agallocha	35
D12	Petafolia A	A. sinensis	30
D13	(+)-8α-Hydroxyeudesma-3,11(13)-dien-14-al	A. sinensis	31
D14	Selina-3,11-dien-9,15-diol	A. sinensis	31
D15	(+)-Eudesma-3,11(13)-dien-8α,9β-diol	A. sinensis	56
D16	(5S,7S,10S)-(-)-Selina-3,11-dien-9-one	A. agallocha	35
D17	Methyl-15-oxo-eudesmane-4,11(13)-dien-12-oate	A. crassna A. malaccensis	57 55
D18	12,15-Dioxo-selina-4,11-dine- [Selina-4,11-diene-12,15-dial]		
D19	Selina-4,11-dien-14-oic acid	A. sinensis A. agallocha	31,32 42
D20	Selina-4,11-dien-14-al	A. agallocha	42
D21	(-)-10-epi-γ-eudesmol	A. malaccensis	33
D22	Eudesma-4-en-8,11-diol	A. crassna	57
		A. malaccensis	55
D23	Eudesma-4-en-11,15-diol	A. sinensis	31
		A. crassna	57
D24	12-hydroxy-4(5),11(13)-eudesmadien-15-al	A. sinensis	31, 30
D25	(7S,8R,10S)-(+)-8,12-dihydroxy-selina-4,11-dien-14-al	A. sinensis	52
D26	(+)-9β-hydroxyeudesma-4,11(13)-dien-12-al	A. sinensis	31
D27	9-hydroxy-selina-4,11-dien-14-oic acid	A. agallocha	42
D28	(7S,9S,10S)-(+)-9-hydroxy-selina-4,11-dien-14-al	A. sinensis	31,52,30
D29	(+)-eudesma-4,11(13)-dien-8α,9β-diol	A. sinensis	31
D30	(+)-eudesma-4(14),11(13)-dien-8α,9β-diol	A. sinensis	31
D31	5-desoxylongilobol	A. sinensis	31
		A.crassna	40
D32	Ent-4(15)-eudesmen-1α,11-diol	A. sinensis	52
D33	Eudesmane-1β,5α,11-triol	A. sinensis	52
D34	(–)-7β-H-eudesmane-4α,11-diol	A. sinensis	52

D35	(4R,5R,7S,9S,10S)-(-)-eudesma-11(13)-en-4,9-diol	A. sinensis	52
D36	Selin-11-en-4a-ol	A. sinensis	31,30
D37	(2R,4aS)-2-(4a-methyl-l,2,3,4,4a,5,6,7-octahydro-2-naphthyl)-pro- pan-2-ol	A. agallocha	43
D38	(S)-4a-methyl-2-(1-methylethy1)-3,4,4a,5,6,7-hexahydronaptha- lene	A. agallocha	43
D39	(S)-4a-methyl-2-(1-methylethylidene)-1,2,3,4,4a,5,6,7-octa- hydronaphthalene	A. agallocha	43
D40	(2R,4aS)-4a-methyl-2-(1-methylethenyl)-l,2,3,4,4a,5,6,7-octa- hydronaphthalene	A. agallocha	43
D41	Dehydrobaimuxinol	A. sinensis	29,47
D42	4-Hydroxyl-baimuxinol	A. sinensis	58
D43	Baimuxinol	A. sinensis	29,47
		A. agallocha	35,45
D44	β-Agarofuran	A. sinensis	29,48,59
D45	Isobaimuxinol	A. sinensis	48
D46	Baimuxifuranic acid	A. sinensis	31,49
D47	(1S,2R,6S,9R)-6,10,10-trimethyl-ll-oxatricyclo[7.2.1.01,6]dodec- ane-2-carbaldehyde	A. agallocha	27
D48	4-hydroxy-dihydro-agarofuran	A. agallocha	46
D49	α-Agarofuran	A. agallocha	45
	-	A. malaccensis	33
D50	Epoxy-β-agarofuran	A. agallocha	27
D51	Dihydro-β-agarofuran	A. agallocha	45
D52	(1R,6S,9R)-6,10,10-trimethyl-11-oxatricyclo[7.2.1.0]dodecane	A. agallocha	43
D53	3,4-dihydroxy-dihydro-agarofuran	A. agallocha	46
D54	Nor-ketoagarofuran	A. agallocha	46
D55	(1R,2R,6S,9R)-6,10,10-trimethyl-11-oxatricyclo[7.2.1.0]dodecan-2- ol	A. agallocha	43
D56	Agalleudesmanol A	A. agallocha	26
D57	Agalleudesmanol B	A. agallocha	26
D58	Agalleudesmanol C	A. agallocha	26
D59	Agalleudesmanol D	A. agallocha	26
D60	Agalleudesmanol E	A. agallocha	26
D61	Agalleudesmanol F	A. agallocha	26
D62	Agalleudesmanol G	A. agallocha	26
D63	Agalleudesmanol H	A. agallocha	26
D64	Agalleudesmanol I	A. agallocha	26
D65	5β,7β-H-elema-1,3-dien-11,13-dihydroxy-11-methyl ester	Aquilaria sp.	50
D66	5 $\beta$ ,7 $\beta$ -H-4 $\alpha$ -hydroxy-eudesma-11,13-dihydroxy-11-methyl ester	Aquilaria sp.	50
D67	5α,7α-H-4(14)-ene-eudesma-11,13- dihydroxy-11-methyl ester	Aquilaria sp.	50

**Table 5:** Eudesmane-type sesquiterpenes from agarwood

#### 2.1.5. E. Eremophilanes (valencanes)

The chemical structures of eremophilane-type sesquiterpenes from agarwood consist two six-membered rings (**E1** to **E38**) are presented in Figure 7, and Table 6. The reported eremophilanes contain a tri-oxygenated isopropyl group (**E6**, **E14, E23, E24, E29, E33**, and **E34**), and an 11-methyl ester functionality (**E24, E29**, and **E34**). The *G. salicifolia* compound, *rel*-4b,5b,7b-eremophil-9-en-12,8a-olide (**E11**) is the only one of an eremophilane containing an 8,12-lactone ring [60]. The *A. agallocha* essential oils compound **E36** exhibits a norskeleton of eremophilane [27], which might be a degradation product of major agarwood compound, dihydrokaranone (**E25**). The eremophilanes, (+)-(4*S*,*5R*)-karanone (**E22**) and (+)-(4*S*,*5R*)-dihydrokaranone (**E25**) are unsaturated and conjugated ketones. These two compounds present in most of the essentials and extracts of *Aquilaria* species, except for *A. malaccensis* from Indonesia, and are characteristic constituents of agarwood [25]. The compounds **E16** and **E26** are a pair of epimers at C-7, and have strong long-lasting pennyroyal-like minty smell [58]. Chemical examination of the ethyl ether extract of *Aquilaria* sp. collected in Thailand, resulted in the isolation and structure determination of eremophilane sesquiterpenes, **E40**, and **E41** [50].



Figure 7: Chemical structures of eremophilane-type sesquiterpenes from agarwood.

No.	Name	Source	Ref.
E1.	Eremophila-9,11(13)-dien-12-ol	A. agallocha	24
E2.	Valenc- or eremophil-9-en-12-al (tentative)	A. agallocha	24
E3.	Jinkoh-eremol	A. malaccensis	28
E4	(1β,3α,4aβ,5β,8aα)-4,4a-dimethyl-6(prop-l-en-2-yl) octahydronaphtha-lene-1,8a(1H)-diol	A. crassna	57
E5	(1aβ,2β,3β,4aβ,5β,8aβ)-octahydro-4a,5-dimeth- yl-3-(1-methylethenyl)-3H-naphth[1,8a-b]oxiren-2-ol	A. malaccensis	55
E6	Eremophil-9-ene-11,12,13-triol	Aquilaria spp.	38
E7	(+)-9β,10β-epoxyeremophila-11(13)-en	A. sinensis	31
	(1β,4aβ,7β,8aβ)-octahydro-7-[1-(hydroxymethyl)	A. malaccensis	55
E8	ethenyl]-1,8a-dimethylnaphthalen-4a(2H)-ol	A. sinensis	31,61
E9	2-[(2β,4aβ,8β,8aβ)-decahydro-4α-hydroxy-8,8a-di- methylnaphthalen-2-yl]prop-2-enal	A. malaccensis A. sinensis	55 31
E10	11,13-dihydroxy-9(10)-ene-8β,12-epoxyemophilane	A. crassna Aquilaria spp.	40 38
E11	rel-4β,5β,7β-eremophil-9-en-12,8α-olide	G. salicifolia	60
E12	8,12-epoxy-eremophila-9,11(13)-diene	A. agallocha	24
E13	Eremophil-9(10)-ene-11,12-diol	G. salicifolia	41
E14	4β,7α-H-eremophil-9(10)-ene-11,12,13-triol	G. salicifolia	41
E15	4β,7α-H-eremophil-9(10)-ene-12,13-diol	G. salicifolia	41
E16	7β-H-9(10)-ene-11,12-epoxy-8-oxoeremophilane	A. sinensis	58
E17	Ligudicin C	A. sinensis	53, 62
E18	()-Eremophila-9-en-8β,11-diol	A. sinensis	31
-	en all and the second	A. crassna	57
E19	4β,7α,8α-H-eremophil-9(10)-ene-8,12-epoxy-11α,13- diol	G. salicifolia	41
E20	Cyclodebneyol	A. sinensis	37
E21	Dehydro-jinkoh-eremol	A. agallocha	42
E22	(+)-(4S,5R)-Karanone	A. agallocha	35
E23	4β,7α-H-eremophil-1(2),9(10)-dien-11,12,13-triol	G. salicifolia	41
E24	4β,7α-H-11,13-dihydroxy-eremophil-1(10)-ene-11- methyl ester	G. salicifolia	41

E25(1) (13,51) diffusional form (11) creation philen-8-one]A. agallocha35,53E26 $7\alpha$ -H-9(10)-ene-11,12-epoxy-8-oxoeremophilaneA. sinensis58,61,62A. crassna40	
E26         7α-H-9(10)-ene-11,12-epoxy-8-oxoeremophilane         A. crassna         40	
A. crassna 40	
E27Petafolia BA. sinensis30	
A. agallocha 42	
E28Neopetasane- [Eremophila-9,11-dien-8-one]A. malaccensis34	
A. sinensis 30,53,58,61,62	
(4S, 5R, 7R) - 11, 12 - dihydroxy-eremophi- la-1(10)-ene-2-oxo-11-methylester       A. crassna       62	
A. malaccensis 24	
E30Kusunol- [Valerianol]A. agallocha25	
A. sinensis 29,61	
<b>E31</b> 2-[(2β,8α,8aα)-8,8a-dimethyl-1,2,3,4,6,7,8,8a-octahy- dronaphthalen-2-yl]propane-1,2-diol <i>A. crassna</i> 57	
E32(+)-trans-NootkatolG. salicifolia41	
2-[(2β,8β,8aα)-8,8a-dimethyl-1,2,3,4,6,7,8,8a-octahy- <b>E33</b> dronaphthalen-2-yl]-3-hydroxy-2-methoxypropanoic <i>A. crassna</i> 57 acid	
E34Methyl crassicidA. crassna63	
E35Valenca-1(10),8-dien-11-olA. agallocha24	
E362,3-dimethyl-r-2-(3-methyl-2-butenyl)-1-cyclohexa- noneA. agallocha27	
E37         11-hydroxy-valenc-1(l0)-en-2-one         A. sinensis         30,31,61	
E38(+)-11-hydroxyvalenc-1(10),8-dien-2-oneA. sinensis31	
E39ValenceneA. malaccensis64	
<b>E40</b> 7β-H-9(10)-ene-emophane-11,13-dihydroxy-11- methyl ester 50	
<b>E41</b> 7α-H-11α,13-dihydroxy-9(10)-ene-8α,12-epoxye- mophane Aquilaria sp. 50	_

**Table 6:** Eremophilane-type sesquiterpenes of agarwood.

#### 2.1.6. F. Guaianes

The sesquiterpene guaianes are structurally coupled with a five- and seven-membered ring structures, and are consisting of a 4,10-dimethyl-7-isopropenyl moiety. The isolated and structure identified guaianes (F1-F47) from the species of Aquilaria and Gyrinops are presented as in Figure 8, and Table 7. The guaianes F2-F11, and F13 bearing a 7-isopropenyl moiety are considered as the characteristic components from the agarwood of A. agallocha, namely kanankoh. The characteristic compound of kanankoh, (-)-guaia-1(10),11dien-15-al (**F7**) has a pleasant  $\beta$ -damascenone-like woody and floral note with a slight cooling side note [35,36]. Among the kanankoh compounds, the isolates F3, F4, F6, F7, F10 and F11 are functionalized at C-14, which is rarely encountered in nature. The compound, (+)-1,5-epoxy-nor-ketoguaiene (F13) is a nor-guaiane with 14 carbons lacking the methyl group at C10. On the other hand, the tricyclic scaffold patchoulenetype compounds F14-F16 are isolated from the agarwood of A. malaccensis [66]. The A. sinensis agarwood compound F19 possesses a 5/6/7 ring system of guaiane skeleton through

C1–C11 linkage. It is interesting to note that the agarwood species A. sinensis is a rich source for various interesting chemical structures. The compounds F17-F31 and F33 are reported from the agarwood of A. sinensis [59]. The guaianefurans (F20-F25) are reported from a agarwood variety of A. sinensis, namely "Lv Qi-Nan" in Chinese [67]. These compounds possess a 5,11-epoxy ring with stereoisomers, and are functionalized at C15 (Figure 8, Table 7). Furthermore, the guaianes F33 and F34, with cleavage of the seven-membered core ring also obtained from the agarwood of A. sinensis [32]. Additionally, the guaianes, F28, F32 and F38, which are possessing a bridge in the seven-membered ring structure are also reported from the agarwood of A. sinensis[61,68]. Among the guaiane sesquiterpenes bearing five-membered lactone, the compounds F35-F37 and F41 are reported from the agarwood of A. filaria and G. salicifolia. These compounds have typical conjugated double bonds within the seven-membered ring, as well as a five-membered  $\alpha,\beta$ -unsaturated lactone [41,68]. Phytochemical examination of A. malaccensis resulted in the isolation of guaiane-type sesquiterpenes F43-F47 [69].



Figure 8: Chemical structures of guaiane-type sesquiterpenes from agarwood.

No.	Name	Source	Ref.
F1.	(+)-Guaia-1(10),11-dien-9-one	A. agallocha	55
F2.	(-)-1,10-epoxyguai-11-ene	A. agallocha	55
F3.	Methyl guaia-1(10),11-diene-15-carboxylate	A. agallocha	35, 55
F4	(-)-Guaia-1(10),11-diene-15-carboxylic acid	A. agallocha	55
F5	α-Bulnesene	A. agallocha	35
F6	(-)-Guaia-1(10),11-dien-15-ol	A. agallocha	55
F7	(-)-Guaia-1(10),11-dien-15-al	A. agallocha	35, 55
F8	α-Guaiene	A. agallocha	35
F9	(-)-Rotundone	A. agallocha	55
F10	(-)-Guaia-1(10),11-dien-15,2-olide	A. agallocha	55
F11	(-)-2α-hydroxyguaia-1(10),11-dien-15-oic acid	A. agallocha	70
F12	(+)-12,13-dihydroxyguaiol	Aquilaria spp.	38
F13	(+)-1,5-epoxy-nor-ketoguaiene	A. agallocha	42
F14	Auranticanol A	A. malaccensis	66
F15	Chamaejasmone D	A. malaccensis	66
F16	Chamaejasmone E	A. malaccensis	66
F17	α-Kessyl alcohol	A. sinensis	71
F18	<i>Epi</i> -guaidiol A	A. sinensis	37
F19	Qinan-guaiane-one	A. sinensis	71
F20	Qinanol E	A. sinensis	67
F21	Qinanol C	A. sinensis	67
F22	Qinanol A	A. sinensis	67
F23	Qinanol B	A. sinensis	67
F24	Qinanol D	A. sinensis	67
F25	Sinenofuranal	A. sinensis	59
F26	Sinenofuranol	A. sinensis	59, 67
F27	1,5;8,12-diepoxyguaia-12-one	A. sinensis	61
F28	3-Oxo-7-hydroxylholosericin A	A. sinensis	61
F29	1α-hydroxy-4α,10α-dimethyl-5βH-octahydro-azulen-8-one	A. sinensis	32
F30	Qinanlactone	A. sinensis	71
F31	7βH-Guaia-1(10)-en-12,8β-olide	A. sinensis	32
F32	1,8-Epoxy-5H-guaia-9-en-12,8-olide	A. filaria	68
F33	1,10-dioxo-4αH-5αH-7βH-11αH-1,10-secoguaia-2(3)-en- 12,8β-olide	A. sinensis	32

F34	1α-hydroxy-4βH-5βH-7βH-11αH-8,9-secoguaia-9(10)-en- 8,12-olide	A. sinensis	32
F35	$2 O x_{0} = 1(10) = 57(11) = 2000 = 12 = 0.0000000000000000000000000000000000$	G. salicifolia	41
F33	2-Oxoguaia-1(10),3,5,7(11),8-pentaen-12,8-olide	A. filaria	68
F36	(4R,5S)-3-Oxo-5,6-dihydro-gweicurculactone	A. filaria	68
F37	(4R)-3-Oxo-gweicurculactone	A. filaria	68
F38	1(5)-Ene-7,10-epoxy-guaia-12-one	A. filaria	68
F39	Guaianolide	G. salicifolia	41
F39	Gualanoide	A. filaria	68
F40	4β,5α,7α,8α-H-3β-hydroxy-1(10)-ene-8,12-epoxy-guaia-12- one	G. salicifolia	41
F41	(–)-Gweicurculactone	G. salicifolia	41
F42	Aromadendrene	A. malaccensis	72
F43	2-Oxo-5β,10β-peroxyl-1αH,4αH,7αH,8βH-guaian-8α,12-olide	A. malaccensis	69
F44	10α-hydroxy-4αH,5αH,7αH,8βH-guaia-1(2)-en-8α,12-olide	A. malaccensis	69
F45	4αH,7αH-14-nor-guaia-1(5)-en-8α,12-olide	A. malaccensis	69
F46	1α,7α-dihydroxy-8oxo-4αH,5αH-guaia-9(10),11(13)-dien-12- oate	A. malaccensis	69
F47	7β,10β-epoxy-4αH-guaia-1(5),11(13)-dien-12-ol	A. malaccensis	69

Table 7: Guaiane-type sesquiterpenes isolated from agarwood species.

## 2.1.7. G. Humulanes

Four humulane-type sesquiterpenes (**G1–G4**) are reported from the agarwood of *A. sinensis* and *A. malaccensis* (Figure 9) [31,66]. The compounds, quilanols A and B (**G1** and **G2**) possess an unprecedented macrocyclic humulene structure with a bicyclic 7/10 ring system [66]. The sesquiterpene  $\beta$ -caryophyllene (**G5**) is reported from from the essential oil of *A. crassna* [73]. Phytochemical examination of *A. malaccensis* resulted in the isolation of humulene-type sesquiterpenes **G6-G9** [69].



Figure 9: Chemical structures of humulane-type sesquiterpenes from agarwood.

#### 2.1.9. H. Prezizaanes

The tricyclic prezizaanetype sesquiterpenes jinkohol II (H1) and jinkohol (H11) are reported from the agarwood

of *A. malaccensis* [25,28,74]. Then the prezizaane-type sesquiterpenes (**H1–H17**), are reported from the agarwood of *Aquilaria* spp. collected in Thailand (Figure 10 and Table 8).





No.	Name	Source	Ref.
Н1.	Jinkohol II	A. malaccensis	28
п.	JITKOHOLII	Aquilaria spp.	75
H2.	Jinkoholic acid	Aquilaria spp.	75
Н3.	Aquilarene E	Aquilaria spp.	76
H4	Aquilarene D	Aquilaria spp.	76
H5	Agarozizanol B	Aquilaria spp.	75
H6	Agarozizanol C	Aquilaria spp.	75
H7	Aquilarene C	Aquilaria spp.	76
H8	Agarozizanol D	Aquilaria spp.	75
H9	Aquilarene B	Aquilaria spp.	76
H10	Aquilarene A	Aquilaria spp.	76
	Balash al	A. malaccensis	74
H11	Jinkohol	Aquilaria spp.	75
H12	Aquilarene F	Aquilaria spp.	76
H13	Aquilarene G	Aquilaria spp.	76
H14	Agarozizanol A	Aquilaria spp.	75
H15	Aquilarene I	Aquilaria spp.	76
H16	Aquilarene H	Aquilaria spp.	76
H17	Aquilarene J	Aquilaria spp.	76

Table 8: Prezizaane-type sesquiterpenes from agarwood.

### 2.1.10. I. Zizaanes

Three tricyclic sesquiterpenes of the zizaane skeleton (I1-I3)

are reported from agarwood of *Aquilaria* spp., collected from Thailand (Figure 11) [75].





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#### 2.1.11. J. Other sesquiterpenoids

In addition to the aforementioned sesquiterpenoids, the species of the agarwood also resulted in the isolation and structure determination different minor sesquiterpenes (Figure 12, Table 9). For example, the eudesmane skeleton compound, 12-hydroxy-dihydrocyperolone (**J1**) is obtained as a new one from the agarwood of *G. salicifolia* [60]. The

daphnauranols B–D (**J2–J4**) exhibiting a rare 5/6/7 ring system were obtained from the agarwood of *A. malaccensis* [66]. Furthermore, the agarwood of *A. malaccensis* also resulted in the isolation and chemical structure identification of tricyclic cadinene-rearranged-sesquiterpenoids with a 6/6/5 ring system, malacinones A and B (**J6** and **J7**) [77]. On the other hand, the compound 1,5,9-trimethyl- 1,5,9-cyclododecatriene (**J5**) is obtained from the from the agarwood of *A. sinensis* [61].



Figure 12: Chemical structures of other sesquiterpenes from agarwood.

No.	Name	Source	Ref.
J1	12-Hydroxy-dihydrocyperolone	G. salicifolia	60
J2	Daphnauranol C	A. malaccensis	66
J3	Daphnauranol B	A. malaccensis	66
J4	Daphnauranol D	A. malaccensis	66
J5	1,5,9-Trimethyl-1,5,9-cyclododeca- triene	A. sinensis	61
J6	Malacinone B	A. malaccensis	77
J7	Malacinone A	A. malaccensis	77

Table 9: Other sesquiterpenes from agarwood.

All these isolation and structure identification reports indicating that agarwood is a rich source for various sesquiterpenes including, acorane, agarospirane, cadinane, eudesmane, eremophilane, guaiane, humulane, prezizaane, or zizaane, etc. Among the reported sesquiterpenes of agarwood, eremophilanes, eudesmanes, and guaianes are widely distributed in various agarwood species. Most of these sesquiterpenes are reported from the agarwood species of *A. agallocha, A. crassna, A. malaccensis*, and *A. sinensis*. Additionally, these sesquiterpenes also reported from the other species of agarwood including *A. filaria*, *G. salicifolia*, and an unidentified *Aquilaria* spp.

#### 2.2. 2-(2-phenylethyl)chromones (PECs)

2-(2-phenylethyl)chromones (PECs) is a member of the class of chromones, which are substituted by a 2-phenylethyl group at C2 position [78]. These compounds has structural resembling with flavonoids, which bears only phenyl group at C-2 position, instead of 2-phenylethyl group present in PECs [78]. PEC derivatives are other major group of constituents in agarwood species [6,12]. The PECs are responsible for the fragrances odor of agarwood burning or heating [12]. The natural PECs are reported from plant species of *Eremophila georgei, Bothriochloa ischaemum* (Gramineae), and agarwood of *Aquilaria* spp [6,12]. Depending on the molecular skeleton, PECs are mainly divided into monomeric 2-(2-phenylethyl)chromone, dimeric 2-(2-phenylethyl)chromones, sesquiterpenoid-4H-chromones and benzylacetone-4H-chromones, and trimeric chromones as described below.

#### 2.2.1. Monomeric 2-(2-phenylethyl)chromone

Following the characteristic structure of the chromone skeleton, monemeric PECs are subdivided into four groups as *Flindersia* type 2-(2-phenylethyl)chromones (FPECs), 5,6,7,8-tetrahydro-2-(2-phenylethyl) chromones (TPECs), mono-epoxy-5,6,7,8-tetrahydro-2-(2-phenylethyl) chromones (EPECs), and diepoxy-5,6,7,8-tetrahydro-2-(2-phenylethyl) chromones (DPECs) (Figure 13).



Figure 13: Chemical structures of monomeric 2-(2-phenylethyl)chromones types of agarwood.

#### 2.2.1a. Flindersia type 2-(2-phenylethyl)chromones (FPECs)

The FPECs are the most abundant PECs in agarwood species (**1–86**). Additionally, a new FPEC (**85a**) is reported from the

MeOH extract of agarwood *Jink* [79]. The chemical structures of FPECs are presented in Figure 14, and their natural source in Table 10.



Figure 14: Skeleton of Flindersia type 2-(2-phenylethyl)chromones from agarwood.

No.	Name	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>	R <sub>5</sub>	R <sub>6</sub>	R <sub>7</sub>	R <sub>8</sub>	Source	Ref.
1	2-[2-(4-hydroxyphe- nyl)ethyl]chromone [Qinanone D]	Н	Н	Н	Н	Н	Н	ОН	Н	A. sinensis	80,81
2	2-[2-(3-hydroxyphe- nyl)ethyl]chromone [Qinanone E]	Н	Н	Н	Н	Н	ОН	Н	Н	A. sinensis	80
3	2-[2-(2-hydroxyphe- nyl)ethyl]chromone [Qinanone F]	Н	Н	Н	Н	ОН	Н	Н	Н	A. sinensis A. malac- censis	80 82
4	8-hy- droxy-2-(2-phenyleth- yl)chromone	Н	Н	н	ОН	Н	Н	н	Н	A. sinensis A. filaria A. malac- censis	83 84 82
5	7-hy- droxy-2-(2-phenyleth- yl)chromone	Н	Н	ОН	Н	Н	Н	Н	Н	A. malac- censis	78
6	6-hy- droxy-2-(2-phenyleth- yl)chromone	Н	ОН	н	Н	Н	Н	Н	н	Kalimantan A. sinensis A. malac- censis A. filaria G. crassna A q u i l a r i a spp.	86 53,80,87,88 34,82 84 89 90
7	5-hy- droxy-2-(2-phenyleth- yl)chromone	ОН	Н	Н	Н	Н	Н	Н	Н	A. sinensis A. malac- censis	91 82
8	(S)-2-(2-hydroxy- 2-phenylethyl) chromone	Н	Н	Н	Н	Н	Н	Н	S-OH	A. crassna A. filaria	89 84
9	(R)-2-(2-hydroxy- 2-phenylethyl) chromone	Н	Н	Н	Н	Н	Н	Н	R-OH	A. crassna A. sinensis A. filaria	89 92,93 84
10	2-(2-phenylethyl) chromone [flidersia- chromone]	Н	н	Н	н	Н	Н	Н	Н	Vietnam A. agallo- cha A. sinensis A. malac- censis A. filaria A q u i l a r i a spp.	94 35 37,62,80,91,92,95 34,82,85 68,84 90
11	7-methoxy-2-(2- phenylethyl)-4H- chromen-4-one	Н	Н	OCH <sub>3</sub>	Н	н	Н	н	Н	A. malac- censis A. sinensis	82,96 53,62,91

12	6-me- thoxy-2-(2-phenyleth- yl)chromone [AH4]	Н	OCH <sub>3</sub>	Н	н	н	н	н	Н	Kalimantan A. sinensis A. agallo- cha A. malac- censis A q u i l a r i a spp.	86 29,95 97 34,82 90
13	2-[2-(4-methoxyphe- nyl)ethyl]chromone	Н	Н	Н	Н	н	Н	OCH <sub>3</sub>	Н	Vietnam A. agallo- cha A. malac- censis A. sinensis	94 35,98 82,96 80,91,99
14	5,8-dihy- droxy-2-(2-phenyleth- yl)chromone [AH <sub>7</sub> ]	ОН	н	Н	ОН	Н	Н	н	Н	Kalinantan A. sinensis	100 91,99,101
15	5-hydroxy-2-[2-(2-hy- droxyphenyl)ethyl] chromone	OH	Н	Н	Н	ОН	Н	Н	Н	A. crassna	102
16	5,6-dihy- droxy-2-(2-phenyleth- yl)chraomone	OH	ОН	Н	Н	н	Н	Н	Н	A. crassna A. malac- censis	103 82
17	6-hydroxy-2-[2-(4-hy- droxyphenyl)ethyl] chromone	Н	ОН	Н	Н	Н	н	ОН	Н	A. malac- censis A. sinensis A. filaria G. salicifolia A. crassna	85 81,87 84 54 102
18	6-hydroxy-2-[2-(2-hy- droxyphenyl)ethyl] chromone	Н	ОН	Н	Н	ОН	Н	Н	Н	A. malac- censis A. sinensis	85 80,92,87
19	6,7-dihydroxy -2-(2-phenylethyl) chromone	Н	ОН	ОН	Н	н	Н	Н	Н	G. salicifolia A q u i l a r i a spp. A. sinensis Jinko	54 104 37 79
20	6,8-dihy- droxy-2-(2-phenyleth- yl)chromone	н	ОН	н	ОН	Н	Н	н	Н	A. malac- censis A. sinensis A. filaria A q u i l a r i a spp.	85 88,92,99 84 104
21	2-[2-hydroxy-2-(4-hy- droxyphenyl)ethyl] chromone	Н	Н	Н	Н	Н	Н	ОН	ОН	A. sinensis	83
22	6-hydroxy-2-(2-hy- droxy-2-phenylethyl) chromone	Н	ОН	Н	Н	Н	Н	Н	ОН	A. sinensis	95, 93
23	6-methoxy-7-hy- droxy-2-(2-phenyleth- yl) chromone	Н	OCH <sub>3</sub>	ОН	Н	Н	Н	Н	Н	A. sinensis	93,105

24	6-hydroxy-5-me- thoxy-2-(2-phenyleth- yl)chromone	OCH <sub>3</sub>	ОН	н	н	Н	Н	н	Н	A. sinensis	62
25	5-hydroxy-6-me- thoxy-2-(2-phenyleth- yl)chromone	ОН	OCH <sub>3</sub>	Н	Н	Н	Н	Н	Н	A. sinensis A. malac- censis	95 82,96
26	6-hydroxy-7-me- thoxy-2-(2-phenyleth- yl)chromone	н	ОН	OCH <sub>3</sub>	н	Н	Н	н	Н	A. malac- censis A. sinensis A. filaria	85 53,62 84
27	6-hydroxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	Н	ОН	Н	н	Н	Н	OCH <sub>3</sub>	н	A. sinensis A. crassna A. filarial A. malac- censis	80,81,88,99 106 84 82
28	6-methoxy-2-[2-(4-hy- droxyphenyl)ethyl] chromone [Aquilar- one H]	Н	OCH <sub>3</sub>	Н	н	Н	Н	ОН	Н	A. sinensis	88,107
29	6-methoxy-8-hy- droxy-2-(2-phenyleth- yl) chromone	Н	OCH <sub>3</sub>	Н	ОН	Н	Н	Н	Н	A. crassna	108
30	6-methoxy-2-[2-(2-hy- droxyphenyl)ethyl] chromone	Н	OCH <sub>3</sub>	Н	н	ОН	Н	Н	Н	A. crassna	103
31	6-methoxy-2-[2-(3-hy- droxyphenyl)ethyl] chromone	Н	OCH <sub>3</sub>	Н	Н	Н	ОН	Н	Н	A. sinensis	88,107
32	2-[2-(3-hydroxy-4-me- thoxyphenyl)ethyl] chromone [Qinanone A]	Н	Н	Н	Н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	80
33	2-[2-(3-methoxy-4-hy- droxyphenyl)ethyl] chromone [Qinanone B]	Н	Н	Н	Н	Н	OCH <sub>3</sub>	ОН	Н	A. sinensis A. crassna	80,81 89
34	7-hydroxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	Н	Н	ОН	Н	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	62
35	2-[2-(2-hydroxy-4-me- thoxyphenyl)ethyl] chromone [Qinanone C]	Н	Н	Н	Н	ОН	Н	OCH <sub>3</sub>	Н	A. sinensis	80
36	7-methoxy-2-[2-(4-hy- droxyphenyl)ethyl] chromone	Н	Н	OCH <sub>3</sub>	Н	Н	Н	ОН	Н	A. sinensis A. crassna	62 109
37	6-hydroxy-8-chloro- 2-(2-phenylethyl) chromone	Н	ОН	Н	Cl	Н	Н	Н	Н	A. sinensis A. filaria A. crassna A. malac- censis	91,93,110 84 63 111

38	2-[2-hydroxy -2-(4-methoxyphenyl) ethyl]chromone	н	н	н	Н	Н	Н	OCH <sub>3</sub>	ОН	A. sinensis A. crassna	90 89
39	5,8-dihy- droxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	ОН	Н	Н	ОН	Н	Н	OCH <sub>3</sub>	н	A. sinensis G. salicifolia	105 111
40	6,7-dime- thoxy-2-(2-phenyleth- yl)chromone [AH6]	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	н	Н	н	Н	Kalinantan A. sinensis A. agallo- cha Kyara 1st (Vietnam) A. malac- censis A. crassna A. filaria A q u i l a r i a	86 53,62,112 88,92,95 83,105 97 36 34,82 106 84 90
41	5,8-dihydroxy-6-me- thoxy-2-(2-phenyleth- yl)chromone	ОН	OCH <sub>3</sub>	Н	ОН	Н	Н	Н	Н	spp. A. sinensis	113
42	6-me- thoxy-2-[2-(3-me- thoxyphenyl)ethyl] chromone [AH5]	Н	OCH <sub>3</sub>	Н	Н	Н	OCH <sub>3</sub>	Н	Н	Kalimantan A. sinensis A. malac- censis	86 53,112,95 82,96
43	2-me- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	н	OCH <sub>3</sub>	н	н	Н	Н	OCH <sub>3</sub>	н	A. agallo- cha A. sinensis A. malac- censis	35,98 29,112,88 96
44	6,8-dihy- droxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	н	ОН	Н	ОН	Н	Н	OCH <sub>3</sub>	н	A. sinensis Aquilaria spp.	99,114 104
45	6-hydroxy-2-[2-(3-hy- droxy-4- methoxyphenyl) ethyl]chromone [Aquilarone l]	н	ОН	Н	н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis Aquilaria spp.	80,81,93,107 90
46	6-hydroxy-7-me- thoxy-2-[2-(4-hydrx- oyphenyl)ethyl] chromone	Н	ОН	OCH <sub>3</sub>	Н	Н	Н	ОН	н	A. sinensis G. salicifolia A q u i l a r i a spp.	93,115 111 116
47	6-hydroxy-2-[2-(3-me- thoxy-4- hydroxyphenyl)ethyl] chromone	н	ОН	Н	Н	Н	OCH <sub>3</sub>	ОН	Н	A. sinensis Aquilaria spp. Aquilaria spp.	80,93,114,117 104 75
48	6,7-dihy- droxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	н	ОН	ОН	н	Н	Н	OCH <sub>3</sub>	н	A. sinensis G. salicifolia A q u i l a r i a spp.	114,115 54 104

49	5-hydroxy-8-me- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	ОН	Н	Н	OCH <sub>3</sub>	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	99
50	5-hydroxy-6-me- thoxy-2-[2-(3-me- thoxyphenyl)ethyl] Chromone	ОН	OCH <sub>3</sub>	Н	Н	Н	OCH <sub>3</sub>	Н	н	A. sinensis	53
51	5-hydroxy-7-me- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	ОН	Н	OCH <sub>3</sub>	Н	Н	н	OCH <sub>3</sub>	Н	A. sinensis	113
52	6-hydroxy-5-me- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	OCH <sub>3</sub>	ОН	Н	Н	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	53
53	6-hydroxy-8-chloro-2- [2-(4-hydroxyphenyl) ethyl]chromone	Н	ОН	Н	Cl	Н	Н	ОН	Н	Aquilaria spp.	116
54	5-Hydroxy-6-me- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	ОН	OCH <sub>3</sub>	Н	Н	Н	Н	OCH <sub>3</sub>	Н	A. malac- censis A. sinensis	96 88
55	6,7-dime- thoxy-2-[2-(3-hy- droxyphenyl)-ethyl] chromone	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	Н	ОН	Н	Н	A. sinensis	81
56	6-methoxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl] chromone	Н	OCH <sub>3</sub>	Н	н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis A. crassna A q u i l a r i a spp.	88 106 90
57	6-methoxy-7-hy- droxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	Н	OCH <sub>3</sub>	ОН	Н	Н	Н	OCH <sub>3</sub>	Н	A. malac- censis A. sinensis A. crassna	34 81,99,114 102,108
58	6-hy- droxy-2-[2-(3,4-dime- thoxyphenyl)ethyl] chromone	Н	ОН	Н	Н	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	Н	A. sinensis	81,114
59	6,7-dime- thoxy-2-[2-(2-hy- droxyphenyl)ethyl] chromone	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	ОН	н	н	Н	A. sinensis	53,92
60	6-methoxy-2-[2-(3- methoxy-4-hydroxy- phenyl)ethyl] chromone	Н	OCH <sub>3</sub>	Н	Н	Н	OCH <sub>3</sub>	ОН	н	A. malac- censis A. sinensis A. crassna A q u i l a r i a spp.	85 53,92,88,105 106 104

61	6-hydroxy-7-me- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	Н	ОН	OCH <sub>3</sub>	н	н	Н	OCH <sub>3</sub>	Н	A. sinensis G. salicifolia A. filaria	114 54 84
62	7-hydroxy-2-[2-(3-me- thoxy-4-hydroxyphe- nyl)ethyl] chromone	Н	Н	OH	Н	Н	OCH <sub>3</sub>	ОН	Н	A. sinensis	81
63	(R)-6,7-dimethoxy-2- (2-hydroxy-2-phenyl- ethyl)chromone	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	Н	Н	Н	Н	R-OH	A. sinensis	93,114
64	(S)-6,7-dimethoxy-2- (2-hydroxy-2-phenyl- ethyl)chromone	н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	н	н	н	S-OH	A. sinensis	93,114
65	6,7-dime- thoxy-2-[2-(4-hy- droxyphenyl)ethyl] chromone [Qinanone G]	н	OCH <sub>3</sub>	OCH <sub>3</sub>	Н	н	н	ОН	н	A. sinensis	81,83,92,114
66	6,7-dime- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone [AH8]	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	Н	н	Н	OCH <sub>3</sub>	Н	Kalinantan A. sinensis A. malac- censis A. crassna	100 29,53,56,62 34 102
67	7-chloro-8-hydroxy-2- [2-(4-methoxyphenyl) ethyl]chromone	Н	Н	CI	ОН	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	53
68	8-chloro-6-hydroxy-2- [2-(4-methoxyphenyl) ethyl]chromone	Н	CI	Н	ОН	Н	Н	OCH <sub>3</sub>	Н	A. sinensis A. crassna	93,110 63,106
69	5,8-dihy- droxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl] chromone	ОН	Н	Н	ОН	Н	ОН	OCH <sub>3</sub>	Н	G. salicifolia	54
70	5,6-dihy- droxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl] chromone	ОН	ОН	Н	Н	н	ОН	OCH <sub>3</sub>	Н	A. sinensis	62
71	5,8-dime- thoxy-2-[2-(3-ace- toxyphenyl)ethyl] chromone	OCH <sub>3</sub>	Н	Н	OCH <sub>3</sub>	Н	OCO- CH <sub>3</sub>	н	Н	A. agallo- cha	97
72	6-me- thoxy-2-[2-(3,4,5-tri- hydroxyphenyl)ethyl] chromone	н	OCH <sub>3</sub>	н	Н	ОН	ОН	ОН	н	A. sinensis	113

73	6,8-dihydrx- oy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl]chromone	Н	ОН	Н	ОН	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	107,115
74	6,8-dihy- droxy-2-[2-(3-me- thoxy-4-hydroxyphe- nyl)ethyl]chromone	Н	ОН	Н	ОН	Н	OCH <sub>3</sub>	ОН	Н	A. sinensis	118
75	8-chloro-6-hydroxy-2- [2-(3-hydroxy-4-me- thoxyphenyl)ethyl] chromone	Н	ОН	Н	CI	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis A. crassna	92,93,114 106
76	2-[2-(4-glucosy- loxy-3-methoxyphe- nyl)ethyl]chromone	Н	Н	н	н	Н	OCH <sub>3</sub>		glu	A. sinensis	119
77	5-Methoxy-6-hy- droxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl]chromone	OCH <sub>3</sub>	ОН	н	Н	Н	ОН	OCH <sub>3</sub>	н	A. sinensis	114
78	6,7-dime- thoxy-2-[2-(3-me- thoxy-4-hydroxylphe- nyl)ethyl]chromone	Н	OCH <sub>3</sub>	OCH <sub>3</sub>	н	Н	OCH <sub>3</sub>	ОН	н	A. sinensis Aquilaria spp.	81,114,115 90
79	6-methoxy-7-hy- droxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl]chromone	Н	OCH <sub>3</sub>	ОН	Н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis G. salicifolia Aquilaria spp.	115 111 75
80	5-hydroxy-6,7-dime- thoxy-2-[2-(4-me- thoxyphenyl)ethyl] chromone	ОН	OCH <sub>3</sub>	OCH <sub>3</sub>	н	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	91
81	6,7-dime- thoxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl]chromone	н	OCH <sub>3</sub>	OCH <sub>3</sub>	Н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis A. crassna	81,114,115 102
82	6-hydroxy-7-me- thoxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl]chromone	Н	ОН	OCH <sub>3</sub>	Н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis G. salicifolia A q u i l a r i a spp. A q u i l a r i a spp.	81,107,114 111 90 75

83	7-hydroxyl-6-methox- yl-2-[2-(4-hydrox- yl-3-methoxylphenyl) ethyl]chromone	Н	OCH <sub>3</sub>	ОН	Н	н	OCH <sub>3</sub>	ОН	Н	A. sinensis	120
84	5-hydroxy-6-me- thoxy-2-[2-(3-hy- droxy-4-methoxyphe- nyl)ethyl]chromone	ОН	OCH <sub>3</sub>	н	Н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis A. crassna	88 102
85	6-hydroxy-7-me- thoxy-2-[2-(3-me- thoxy-4-hydroxyphe- nyl)ethyl]chromone [Aquilarone G]	Н	ОН	OCH <sub>3</sub>	н	н	OCH <sub>3</sub>	ОН	Н	A. sinensis Aquilaria spp.	107 90
85a	6,7-dihy- droxy-2-[2-(3'-hy- droxy-4'-methoxy- phenyl)ethyl] chromone	-	-	-	-	-	-	-	-	Jinko	79
86	6-me- thoxy-2-[2-(3,4,5-tri- hydroxyphenyl)ethyl] chromone	н	OCH <sub>3</sub>	н	Н	ОН	ОН	ОН	Н	A. sinensis	113

Table 10: Flindersia type 2-(2-phenylethyl)chromones from agarwood species.

The commonly observed substituents in FPECs core structure are hydroxy and methoxy groups, and are substituted at C-6, followed by C-7, C-5 and C-8. The methoxy functional groups appear more frequently at C-7 than hydroxyl groups. However, it is interesting to note that five chlorinated FPECs (**37**, **53**, **67**, **68** and **75**), are reported from the agarwood species (Figure 14, Table 10). It is also reported the only glycosylated FPEC (**76**) from *A. sinensis*. The only acetyl FPEC (**71**) reported from the agarwood of *A. agallocha*.

# 2.2.1b. 5,6,7,8-tetrahydro-2-(2-phenylethyl)chromones (TPECs)

The isolated and structure identified highly oxidized TPECs (**87–135**) from agarwood species are presented as Figure 15, and Table 11. Further, chemical examination of whole-tree agarwood-inducing technique (Agar-Wit) from 8 years old A. sinensis, resulted in the isolation of TPEC compounds **135a**, **135b**, and **135c** (Figure 14) [121].



Figure 15: General chemical structure of TPECs from agarwood species.

No.	Name	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>	R <sub>5</sub>	R <sub>6</sub>	<b>R</b> <sub>7</sub>	R <sub>8</sub>	Source	Ref.
87	(6S,7S,8S)-6,7,8-trihydroxyl-2-(3-hy- droxyl-4-methoxyphenylethyl)-5,6,7,8- tetrahydro-4H-chromen-4-one.	Н	a-OH	<i>а</i> -ОН	<i>a</i> -OH	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	120
88	(6S,7S,8S)-6,7,8-trihydroxyl-2-(4-hy- droxyl-3-methoxyphenylethyl)-5,6,7,8- tetrahydro-4H-chromen-4-one	н	α-ОН	а-ОН	<i>α</i> -OH	Н	OCH <sub>3</sub>	ОН	Н	A. sinensis	120
89	6,7-dihydroxy-5,6,7,8-tetrahy- dro-2-(2-(4-methoxy phenyl)ethyl) chromone	Н	a-OH	<i>α</i> -OH	Н	Н	Н	OCH <sub>3</sub>	Н	A. crassna	122
90	6,7-dihydroxy-2-(2-phenyleth- yl)-5,6,7,8-tetrahydrochromone	Н	α-OH	<i>а</i> -ОН	Н	Н	Н	Н	Н	A. sinensis Aquilaria spp.	95 90
91	(6S,7S,8R)-6,7-dihydroxy-8-chloro- 5,6,7,8-tetrahydro-2-(2-(3-hydroxy-4- methoxyphenyl)ethyl)chromone	н	a-OH	a-OH	β-Cl	Н	ОН	OCH <sub>3</sub>	Н	A. crassna	122
92	(5S,6R,7R)-5,6,7-trihydroxy-2-(3-hy- droxy-4-methoxyphenylethyl)-5,6,7,8- tetrahydro-4H-chromen-4-one	а-ОН	<i>α</i> -OH	<i>α</i> -OH	Н	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	91,123
93	rel-(5R,6S,7R)-5,6,7,8-tetrahydro-5,6,7- trihydroxy-2-[2-(4-methoxyphenyl) ethyl]-4H-1-benzopyran-4-one	а-ОН	α-ОН	<i>β</i> -OH	Н	Н	Н	OCH <sub>3</sub>	Н	A. malaccensis	34
94	(6R,7S,8S)-6,7,8-trihydroxy-2-(4-hy- droxyl-3-methoxyphenylethyl)-5,6,7,8- tetrahydro-4H-chromen-4-one.	<i>β</i> -OH	<i>β</i> -OH	<i>β</i> -OH	Н	Н	OCH <sub>3</sub>	ОН	Н	A. sinensis	91

95	rel-(5R,6S,7R)-5,6,7,8-tetrahydro-5,6,7- trihydroxy-2-(2-phenylethyl)-4H-1- benzopyran-4-one	а-ОН	α-ОН	<i>β</i> -OH	н	Н	Н	Н	Н	A. malaccensis	34
96	(5S,6S,7R)-5,6,7-trihydroxy-2-[2-(hy- droxylphenyl)ethyl]-5,6,7,8-tetrahydro- chromone [AH9]	<i>α</i> -OH	<i>β</i> -OH	α-ОН	Н	ОН	Н	Н	Н	Kalimantan	100
97	(5S,6R,7S)-5,6,7-trihydroxy-2-(3-hy- droxy-4-methoxyphenylethyl)-5,6,7,8- tetrahydro-4H-chromen-4-one	<i>α</i> -OH	α-ОН	<i>β</i> -OH	Н	Н	ОН	OCH <sub>3</sub>	Н	A. malaccensis	34
98	Agarotetrol [AH1]	α-ΟΗ	<i>β</i> -OH	<i>β</i> -OH	a-OH	Н	Н	Н	Н	A. agallocha Kalimantan Aquilaria spp. A. sinensis	124 125 104 126, 127
99	Aquilarone B	a-OH	a-OH	α-ОН	β-ΟΗ	Н	Н	н	Н	A. sinensis Aquilaria spp.	107,127 90
100	Tetrahydrochromone B	$\beta$ -OCH $_3$	a-OH	a-OH	β-ΟΗ	Н	Н	Н	Н	A. sinensis	127
101	5α,6β,7β-trihydroxy-8α-me- thoxy-2-(2-phenylethyl)-5,6,7,8-tetra- hydrochromone [AH17]	а-ОН	<i>β</i> -OH	β-OH	a-OCH <sub>3</sub>	Н	Н	Н	Н	Kalimantan A. sinensis	128 127
102	(5R,6R,7S,8R)-2-(2-phenylethyl)-tetra- hydroxy-5,6,7,8-tetrahydrochromone [AH16]	<i>β</i> -OH	<i>β</i> -OH	а-ОН	<i>β</i> -OH	Н	Н	Н	Н	Kalimantan A. sinensis	129 126
103	Isoagarotetrol [AH2]	<i>α</i> -OH	<i>β</i> -OH	а-ОН	β-ΟΗ	Н	Н	н	Н	Kalimantan A. sinensis Aquilaria spp.	125 95 90

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104	(5R,6S,7S,8R)-2-[2-(4-methoxyphenyl) ethyl]-5,6,7,8-tetrahydroxy-5,6,7,8-tet- rahydrochromone	β-ΟΗ	a-OH	a-OH	<i>β</i> -OH	Н	Н	OCH <sub>3</sub>	Н	Aquilaria spp.	90
105	5α,6β,7α,8β-tetrahydroxy-2-[2-(4-me- thoxyphenyl)ethyl]-5,6,7,8-tetrahydro- chromone [AH2a]	a-OH	<i>β</i> -OH	α-ОН	<i>β</i> -OH	Н	Н	OCH <sub>3</sub>	Н	Kalimantan A. sinensis Aquilaria spp.	130 37,87 90
106	(5S,6R,7S,8R,7'R)-7'-hydroxyisoagaro- tetrol	a-OH	β-ΟΗ	α-ОН	β-ΟΗ	Н	Н	н	R-OH	Kalimantan	131
107	8-chloro-2-(2-phenylethyl)-5,6,7-trihy- droxy-5,6,7,8-tetrahydrochromone	α-ΟΗ	a-OH	а-ОН	β-Cl	Н	Н	Н	Н	A. sinensis Aquilaria spp.	53,95 90
108	5α,6β,7α,8β-tetrahydroxy-2-[2-(2-hy- droxyphenyl)ethyl]-5,6,7,8-tetrahydro- chromone [AH2b]	a-OH	β-ΟΗ	a-OH	<i>β</i> -OH	ОН	Н	Н	н	Kalimantan A. sinensis Aquilaria spp.	130 56 90
109	5α,6β,7β,8α-tetrahydroxy-2-[2-(2-hy- droxyphenyl)ethyl]-5,6,7,8-tetrahydro- chromone (AH <sub>23</sub> )	α-ΟΗ	<i>β</i> -OH	β-ОН	α-OH	ОН	Н	Н	н	Kalimantan	128
110	5α,6β,7β,8α-tetrahydroxy-2-[2-(4-me- thoxyphenyl)ethyl]-5,6,7,8-tetrahydro- chromone[AH1A] [4'-methoxy-agaro- tetrol]	<i>a</i> -OH	<i>β</i> -OH	<i>β</i> -OH	<i>α</i> -OH	Н	Н	OCH <sub>3</sub>	н	Kalimantan A. sinensis Aquilaria spp.	130 127,132 116
111	Aquilarone C	a-OH	α-OH	а-ОН	β-ΟΗ	Н	Н	OCH <sub>3</sub>	Н	A. sinensis Aquilaria spp.	91,107 90
112	(5S,6S,7S,8S)-8-chloro-5,6,7- trihydroxy-2-(phenylethyl)-5,6,7,8- tetrahydrochromone	<i>α</i> -OH	а-ОН	а-ОН	a-Cl	Н	Н	Н	н	A. sinensis	91

113	(5S,6R,7S,8R,7'S)-7'-hydroxyisoagaro- tetrol	α-OH	β-ΟΗ	<i>α</i> -OH	<i>β</i> -OH	Н	Н	Н	S-OH	Kalimantan	131
114	Aquilarone F	<i>α</i> -OH	β-ΟΗ	β-ΟΗ	а-ОН	Н	Н	ОН	Н	A. sinensis Aquilaria spp.	107 90
115	(5R,6S,7S,8R)-2-[2-(4-hydroxy-3- methoxyphenyl)ethyl]-5,6,7,8-tetrahy- droxy-5,6,7,8-tetrahydrochromone	<i>β</i> -OH	α-ΟΗ	<i>α</i> -OH	β-OH	Н	OCH <sub>3</sub>	ОН	н	Aquilaria spp.	90
116	Tetrahydrochromone G	$\beta$ -OCH $_{_3}$	β-ΟΗ	β-ΟΗ	a-OH	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	127
117	Aquilarone E	a-OH	β-ΟΗ	β-ΟΗ	α-ΟΗ	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	107, 127
118	(5R,6S,7S,8R)-2-[2-(3-hydroxy-4-me- thoxyphenyl)ethyl]-5,6,7,8-tetrahy- droxy-5,6,7,8-tetrahydrochromone	β-ОН	α-ΟΗ	<i>α</i> -OH	β-OH	н	ОН	OCH <sub>3</sub>	Н	Aquilaria spp.	90
119	Tetrahydrochromone F	a-OCH <sub>3</sub>	<i>α</i> -OH	<i>α</i> -OH	<i>β</i> -OH	Н	Н	OCH <sub>3</sub>	Н	A. sinensis A. crassna	127 102
120	(5R,6S,7S,8R)-5,6,7-trihydroxy-8-me- thoxy-5,6,7,8-tetrahydro-2-(2-(4-me- thoxyphenyl)ethyl)chromone	β-ОН	a-OH	a-OH	$\beta$ -OCH <sub>3</sub>	Н	Н	OCH <sub>3</sub>	Н	A. crassna	122
121	Tetrahydrochromone E	a-OH	β-ΟΗ	β-ΟΗ	$\alpha$ -OCH <sub>3</sub>	Н	Н	$\operatorname{OCH}_3$	Н	A. sinensis	127
122	5,6,7,8-tetrahydroxy-2-(3-hy- droxy-4-methoxy phenethyl)-5,6,7,8- tetrahydro-4H-chromen-4-one	a-OH	<i>β</i> -OH	β-ΟΗ	α-ΟΗ	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	133
123	Aquilarone D	a-OH	β-ΟΗ	α-ОН	β-ОН	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis Aquilaria spp.	56,107 90

124	Aquilarone A	a-OH	а-ОН	а-ОН	β-ΟΗ	Н	ОН	OCH <sub>3</sub>	н	A. sinensis Aquilaria spp.	107,127 90
125	Tetrahydrochromone A	a-OCH <sub>3</sub>	β-ΟΗ	β-ΟΗ	a-OH	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	127
126	(5R,6R,7R,8R)-8-chloro-5,6,7- trihydroxy-2-(4-methoxyphenethyl)- 5,6,7,8-tetrahydrochromone	<i>β</i> -OH	<i>β</i> -OH	<i>β-</i> ΟΗ	β-Cl	Н	н	OCH <sub>3</sub>	н	A. sinensis	91
127	rel-(5R,6S,7S,8R)-8-chloro5,6,7,8-tet- rahydro-5,6,7-trihydroxy-2-[2-(4-me- thoxyphenyl)ethyl]-4H-1-benzopyran- 4-one	α-ОН	β-OH	β-OH	a-Cl	Н	Н	OCH <sub>3</sub>	н	A. malaccensis A. sinensis	34 127
128	(5R,6R,7R,8S)-8-chloro-5,6,7- trihydroxy-2-(4-methoxyphenethyl)- 5,6,7,8-tetrahydrochromone	<i>β</i> -OH	β-ОН	<i>β</i> -0Η	a-Cl	Н	Н	OCH <sub>3</sub>	н	A. sinensis Aquilaria sp	91 75
129	Tetrahydrochromone I	a-OCH <sub>3</sub>	α-ОН	α-ОН	β-Cl	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	127
130	Tetrahydrochromone D	a-OCH <sub>3</sub>	<i>β</i> -OH	β-ΟΗ	a-Cl	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	127
131	Tetrahydrochromone C	a-OCH <sub>3</sub>	β-ΟΗ	β-ΟΗ	а-ОН	Н	ОН	OCH <sub>3</sub>		A. sinensis	127
132	Tetrahydrochromone H	a-OCH <sub>3</sub>	a-OH	a-OH	β-ΟΗ	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	127
133	Tetrahydrochromone J	a-OCH <sub>3</sub>	а-ОН	а-ОН	β-Cl	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	127
134	8-chloro-5,6,7-trihydroxy-2-(3-hydroxy- 4-methoxyphenethyl)-5,6,7,8-tetrahy- dro-4H-chromon-one	<i>α</i> -OH	α-ΟΗ	а-ОН	β-Cl	Н	ОН	OCH <sub>3</sub>	Н	A. sinensis	134

135	rel-(5R,6S,7S,8R)-8-chloro-5,6,7,8-tet- rahydro-5,6,7-trihydroxy-2-[2-(3-hy- droxy-4-methoxyphenyl)ethyl]- 4H-1-benzopyran-4-one	α-ΟΗ	<i>β</i> -OH	<i>β</i> -OH	a-Cl	Н	ОН	OCH <sub>3</sub>	н	A. malaccensis A. sinensis	34 37
135a	(5S,6R,7S,8S)-8-chloro-5,6,7-trihydroxy- 2-[2-(4'-methoxyphenylethyl)]-5,6,7,8- tetrahydrochromone	а-ОН	<i>β</i> -OH	а-ОН	α-Cl	Н	Н	OCH <sub>3</sub>	Н	A. sinensis	121
135b	(5S,6R,7S,8S)-8-chloro-5,6,7-trihydroxy- 2-(2-phenylethyl)-5,6,7,8- tetrahydrochromone	α-OH	<i>β</i> -OH	а-ОН	α-Cl	Н	Н	н	Н	A. sinensis	121
135c	(5S,6R,7R,8S)-8-chloro-5-ethoxy-6,7-di- hydroxy-2-[2-(3'-hydroxy-4'-methoxy- phenylethyl)-5,6,7,8-tetrahydrochro- mone	α-OCH <sub>2</sub> CH <sub>3</sub>	<i>β</i> -OH	<i>β</i> -OH	α-Cl	Н	ОН	OCH <sub>3</sub>	н	A. sinensis	121

Table 11: 5,6,7,8-tetrahydro-2-(2-phenylethyl)chromones (TPECs) reported from agarwood.

Among the reported TPECs, the compound, agarotetrol (**98**) is a common metabolite found in different samples of agarwood. The Chinese Pharmacopeia (2015 edition), identified agarotetrol (**98**) as a marker compound of agarwood, and standardized the contents needs to be higher than 0.10%. Most of the reported TPECs are oxidized at C-5, C-6, C-7 and C-8 positions, either with hydroxy or methoxy functional groups. It is observed that the reported TPECs at C-6 and C-7 are usually substituted with hydroxyl groups, while the methoxy group at C-5 or C-8. Some of the reported TPECs contains a chlorine substituent at the C-8 position. It is interesting to note that the TPECs **106** and **113** are ethoxy derivatives.

#### 2.2.1c. Mono-epoxy-5,6,7,8-tetrahydro-2-(2-phenylethyl)chromones (EPECs)

Various epoxy-substituted PECs are reported from agarwood species (Figure 16, Table 12). The epoxy group is usually located either at C-5 and C-6, or at C-7 and C-8. However, the EPEC **141** carries an epoxy group located at C-6 and C-7 [135]. Similar to the structures of TPECs, the C-5, C-6, C-7 and C-8 positions of EPECs are oxidized and carry hydroxy or methoxy or epoxy groups, while the methoxy groups usually located at C-5 or C-8. In this connection, the EPECs **136–138** are reported from *A. malaccensis* [34], whereas **136–144**, and **148–150** are reported from the agarwood of *A. sinensis* (Table 12). The EPECs **136** and **144** are obtained from the agarwood of *A. crassna* [108].
# 2.2.1d. Diepoxy-5,6,7,8-tetrahydro-2-(2-phenylethyl) chromones (DPECs)

the species of agarwood (Figure 16, Table 12). The DPECs (**145–147**) are obtained from the agarwood of *A. crassna*, *A. malaccensis* and *A. sinensis* (Table 12).

Various DPECs compounds (145-151), are reported from



Figure 16: Chemical structures of EPECs and DPECs of agarwood.

No.	Name	Source	Ref.
		A. malaccensis	34
136	<i>rel-(1aR,2R,3R,7bS)</i> -1a,2,3,7b-tetrahydro-2,3-dihydroxy-5-[2-(4-methoxyphenyl)ethyl]-7H-ox- ireno[f] [1]benzopyran-7-one	A. sinensis	91
		A. crassna	108
137	<i>rel-(1aR,2R,3R,7bS</i> )-1a,2,3,7b-tetrahydro-2,3-dihydroxy-5-[2-(3-hydroxy-4-methoxyphenyl)	A. malaccensis	34
	ethyl]-7H-oxireno[f][1]benzopyran-7-one	A. sinensis	88
	<i>rel-(1aR,2R,3R,7bS)</i> -1a,2,3,7b-Tetrahydro-2,3-dihydroxy-5-(2-phenylethyl)-7H-oxireno[f] [1] benzopyran-7-one	A. malaccensis	34,82
138		A. sinensis	53,8
139	5,6-epoxy-7β-hydroxy-8β-methoxy-2-(2-phenylethyl)chromone	A. sinensis	88
140	5α,6α-Epoxy-7β,8α,3'-trihydroxy-4'-methoxy-2-(2-phenylethyl)chromone	A. sinensis	113
141	(55,6R,7S,8S)-2-[2-(4'-methoxyphenyl)ethyl]-6,7-epoxy-5,8-dihydroxy-5,6,7,8-tetrahydrochro- mone	A. sinensis	135

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142	Tetrahydrochromone M	A. sinensis	127
143	Tetrahydrochromone L	A. sinensis	127
144	(5R,6S,7S,8S)-2-[2-(4'-methoxyphenyl)ethyl]-7,8-epoxy-5-methoxy-6-hydroxy-5,6,7,8-tetrahy-	A. sinensis	135
144	drochromone	A. crassna	108
		A. crassna	136
145	Oxidoagarochromone B	A. malaccensis	34
		A. sinensis	53,88
		A. crassna	136
146	Oxidoagarochromone A	A. malaccensis	34
		A. sinensis	53,88
		A. crassna	136
147	Oxidoagarochromone C	A. malaccensis	34
147		A. sinensis	127
		A. SITIETISIS	127
148	<i>(55,65,75,85)</i> -2-[2-(3'-hydroxy-4'-methoxyphenyl)ethyl]-7,8-epoxy-5-methoxy-6-hy- droxy-5,6,7,8- tetrahydrochromone	A. sinensis	135
149	( <i>55,65,75,85</i> )-2-[2-(4'-methoxyphenyl)ethyl]-7,8-epoxy-5-methoxy-6-hydroxy-5,6,7,8-tetrahy-	A. sinensis	135
149	drochromone	A. SINENSIS	133
150	(55,65,75,85)-2-[2-(4'-methoxyphenyl)ethyl]-7,8-epoxy-5,6-dihydroxy-5,6,7,8-tetrahydrochro- mone	A. sinensis	135
151	Tetrahydrochromone K	A. sinensis	127
		71. 3111011313	127

Table. 12: EPECs and DPECs from agarwood species.

The C-4' position of the phenylethyl moiety in both EPECs and DPECs are without substitution, or substituted with a methoxy group (Figure 16). Alternatively, both EPECs and DPECs are substituted at C-3' with a hydroxyl and/or C-4' with a methoxyl group (Figure 16).

# 2.2.1e. Other PECs

From the agarwood species of *A. crassna, A. filaria, A. sinensis,* and *G. salicifolia*, seven 2-(2-phenylethenyl)chromones (PEECs) are reported (Figure 17). The compounds PEECs are possess a styryl moiety instead of a phenylethyl moiety at C-2 of chromones. The chemical structures and names are presented as in Figure 17 and Table 13.



Figure 17: Chemical structures of 2-(2-phenylethenyl)chromones (PEECs) from agarwood.

No.	Name	Source	Ref.
152	(E)-2-[2-(3-methoxy-4-hydroxyphenyl)ethenyl]chromone	A. crassna	109
153	(E)-6-methoxy-2-[2-(4-hydroxyphenyl)ethenyl]chromone	A. crassna	109
154	5-hydroxy-2-[2-(4-methoxyphenyl)ethenyl]chromone	G. salicifolia A. filaria	54 68
155	(E)-6-methoxy-2-[2-(3-methoxy-4-hydroxyphenyl)ethenyl] chromone	A. crassna	109
156	6-hydroxy-2-[2-(3-methoxy-4-hydroxyphenyl)ethenyl]chromone	A. sinensis Aquilaria spp.	37,115 116
157	5-hydroxy-2-[2-(3-hydroxy-4-methoxyphenyl)ethenyl]chromone	G. salicifolia	137
158	6,7-dimethoxy-2-[2-(4-hydroxyphenyl)ethenyl]-4H-chromen-4-one	A. sinensis	92

Table 13: 2-(2-phenylethenyl)chromones (PEECs) from agarwood.

### 2.2.2. Dimeric 2-(2-phenylethyl)chromones (DIPECs)

The agarwood species of *A. sinensis* and *A. crassna* are rich source for the compounds DIPECs (Figure 18, Table 14). The DIPECs are isolated and purified by silica gel column chromatography and semi-preparetive HPLC and so on. The chemical structures of DIPECs are identified by spectroscopic data. The single C–C bond linked DIPECs (**159–162**), which are composed of two FPECs (unit A and unit B) through a C5–C5' linkage, are reported from the agarwood "Jinko" from Kalimantan (Figure 18, Table 14). Further, chemical examination of whole-tree agarwood-inducing technique (Agar-Wit) from 8 years old *A. sinensis*, resulted in the isolation of a single C–C bond linked DIPEC, aqulisinone A (**162a**, Figure 18) [121]. The reported remaining DIPECs are linked with a C–O–C bond (Table 14). Most of these C–O–C bond linked DIPECs are composed of a TPEC unit (unit A) and a FPEC unit (unit B) (Figure 18, Table 14). In the DIPECs 167–187, the linkage position of unit A is situated at C-8, while the linkage position of unit B is usually at C-7' or at C-6', except for dimer 176 (Figure 18). The DIPECs 195 and 196 are composed of two EPEC units, which are connected through a C5–O–C6' linkage (Figure 18). It is observed that the DIPECs unit A usually linked at C5 or C8. The reason might be that the conjugated C5 and C8 of 4H-pyran-4-one yielded the stable intermediate carbocations as compared with C6 and C7. The DIPECs AH<sub>10</sub>-AH<sub>15</sub>, and AH<sub>21</sub> (164-166, 168, and 169) are reported from the agarwood "Jinko" from Kalimantan (Table 15). Further,  $AH_{10}$  (**166**) and  $AH_{14}$  (**165**) are also reported from the withered wood of A. sinensis grown in Taiwan [95]. A recent study reported the new DIPECs 198a-198d from the MeOH extract of agarwood Jinko [79]. The DIPEC compounds, aquilasinenones L and M (198e and 198f) are reported from the artificial agarwood originating from A. sinensis [138].



Figure 18: Chemical structures of dimeric 2-(2-phenylethyl)chromones from agarwood.

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No.	Name	Source	Ref.
159	Aquisinenone O	A. sinensis	139
160	2,2'-di-(2-phenylethyl)-8,6'-dihydroxy-5,5'-bichromone [AH <sub>11</sub> ]	Kalimantan A. sinensis	140 139
161	7,4'-dimethoxyaquisinenone O	A. sinensis	139
162	Crassin A	A. crassna A. sinensis	141 139
162a	Aqulisinone A	A. sinensis	121

 Table 14: Dimeric 2-(2-phenylethyl)chromones with C–C bond of agarwood.

No.	Name	Source	Ref.
163	Aquilasinenone K	A. sinensis	142
164	(5S,6S,7R,8S)-2-(2-phenylethyl)-6,7,8-trihydroxy-5,6,7,8-tetrahydro-5-[2-(2-phenyl- ethyl)-7-hydroxy-chromonyl-6-oxy]chromone [AH <sub>15</sub> ]	Kalimantan	143
165	(5S,6R,7S,8S)-2-(2-phenylethyl)-6,7,8-trihydroxy-5,6,7,8-tetrahydro-5-[2-(2-phenyl- ethy)chromonyl-6-oxy]chromone [AH14]	Kalimantan	144
166	(5S,6S,7R,8S)-2-(2-phenylethyl)-6,7,8-trihydroxy-5,6,7,8-tetrahydro-5-[2-(2-phenyl-	A. sinensis Kalimantan	39,95 140
	ethy)chromonyl-6-oxy]chromone [AH10]	A. sinensis	95
167	(5R,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-[2-(2-phenyl- ethyl)chromonyl-6-oxy]chromone	A. sinensis	145
168	(5S,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-[2-(2-phenyl- ethy)chromonyl-6-oxy]chromone [AH13]	Kalimantan	144
169	(5R,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-[2-(2-phenyl- ethy)-7-methoxychromonyl-6-oxy]chromone [AH12]	Kalimantan	144
170	Aquisinenone N	A. sinensis	139
171	(5S,6R,7S,8R)-2-[2-(4-methoxyphenyl)ethyl]-5,6,7-trihydroxy5,6,7,8-tetrahy- dro-8-{2-[2-(4'''-methoxyphenyl)ethyl]chromonyl-6-oxy}chromone	A. sinensis	145
172	(5S,6R,7S,8R)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-[2-(2-phenyl- ethyl)chromonyl-6-oxy]chromone	A. sinensis	145
173	Aquilasinenone J	A. sinensis	142
174	Aquisinenone M	A. sinensis	139
75	Crassin D	A. sinensis	139

176	Crassin B	A. sinensis	141
177	Aquilasinenone C	A. sinensis	142
178	Aquilasinenone B	A. sinensis	142
179	Aquilasinenone E	A. sinensis	142
180	Aquilasinenone D	A. sinensis	142
181	Aquilasinenone A	A. sinensis	142
182	Crassin C	A. crassna	141
183	Aquilasinenone H	A. sinensis	142
184	Aquilasinenone G	A. sinensis	142
185	Aquilasinenone I	A. sinensis	142
186	(5S,6R,7S,8R)-2-[2-(4-methoxyphenyl)ethyl]-5,6,7-trihydroxy-5,6,7,8-tetrahydro- 8-{6-methoxy-2-[2-(3'''-methoxy-4'''-hydroxyphenyl)ethyl]chromonyl-7-oxy} chromone	A. sinensis	145
187	Aquilasinenone F	A. sinensis	142
188	Aquisinenone H	A. sinensis	139
189	4'-methoxyaquisinenone	A. sinensis	139
190	4',7",4"'-trimethoxyaquisinenone l	A. sinensis	139
191	Aquisinenone I	A. sinensis	139
192	7"-methoxyaquisinenone l	A. sinensis	139
193	4',7"-dimethoxyaquisinenone l	A. sinensis	139
194	Aquisinenone L	A. sinensis	139
195	4',4'''-dimethoxyaquisinenone K	A. sinensis	139
196	Aquisinenone K	A. sinensis	139

197	Aquisinenone J	A. sinensis	139
198	4'-methoxyaquisinenone J	A. sinensis	139
198a	(5S,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-[6'-hydroxy- 2-(2-phenylethyl)chromonyl-7'-oxy]chromone [diaquilariachrome A]	Jinko	79
198b	(5S,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-[6'-hydroxy- 2-(2-phenylethyl)chromonyl-8'-oxy]chromone [diaquilariachrome B]	Jinko	79
198c	(5S,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-(3-phenyl- propionyloxy)chromone	Jinko	79
198d	(5S,6R,7R,8S)-2-(2-phenylethyl)-5,6,7-trihydroxy-5,6,7,8-tetrahydro-8-{6'-hydroxy- 2-[2-(4'''-methoxyphenyl)ethyl]chromonyl-7'-oxy}chromone [diaquilariachrome C]	Jinko	79
198e	Aquilasinenones L	A. sinensis	138
198f	Aquilasinenones M	A. sinensis	138

 Table 15: Dimeric 2-(2-phenylethyl)chromones with C–O–C bond of agarwood.

Additionally, double linked 2-(2-phenethyl)chromone dimers (DLPECs) also reported from the species of agarwood. The reported compounds chemical structures (Figure 19) are presented in Table 16. These compounds are composed of a TPEC unit (unit A) and an FPEC unit (unit B) (Figure 19). In most of the DLPECs, the linkage position of unit A is usually at C5 and C7, while the same in unit B is C6' and C7', might be duce to the 6,7-dihydroxy-FPECs provides two adjacent hydroxyl groups to form the two C–O–C bonds (Figure 19). In the TPEC unit, the C–O–C bond linked at C7, while the same for the C–C bond is at C5. On the other hand, in the FPEC unit, the C–C bond linked at C5', C7' or C8' of the chromone moiety (**205–215**), or at C2'',

C3<sup>III</sup> or C4<sup>III</sup> of the phenylethyl moiety (**216–224**) (Figure 19, Table 16). Among these DLPECs, six compounds (199–204) are linked through two C–O–C bonds to form a seven or sixmembered oxygen-carrying heterocyclic ring (Figure 19, Table 16). Six new DLPECs (204a–204f) are reported from the EtOAc extract of artificial agarwood originating from *A. sinensis* [146]. On the other hand, the DLPECs 205–224 contains an unusual 3,4-dihydro-2H-pyran ring connected to two PEC monomeric moieties through a C–O–C bond and a C–C bond (Figure 19, Table 17). Three new C–O–C bond DLPECs, crassin I– K (**224a–224c**) (Figure 19, Table 17), reported from the artificial holing agarwood originating from *A. sinensis* [147].

No.	Name	Source	Ref.
199	Crassin E	A. crassna	148
200	Crassin F	A. crassna	148
201	Crassin G	A. crassna	148
202	AH <sub>21</sub>	Kalimantan	149
203	(+)-4'-Methoxyaquisinenone G	A. sinensis	150

204	(–)-Aquisinenone G	A. sinensis	150
204a	(–)-3'''-hydroxy-4'''-methoxy-aquisinenone G	A. sinensis	146
204b	(+)-3'''-hydroxy-4'',4'''-dimethoxy-aquisinenone G	A. sinensis	146
204c	(+)-3"-hydroxy-4",4"'-dimethoxy-aquisinenone G	A. sinensis	146
204d	(+)-4"'-hydroxy-4",3"'-dimethoxy-aquisinenone G	A. sinensis	146
204e	3"'-hydroxy-4"-demethoxy-crassin G	A. sinensis	146
204f	3'''-hydroxy-crassin G	A. sinensis	146

 Table 16: Double linked 2-(2-phenylethyl)chromones with double C–O–C bonds.



Figure 19: Chemical structures of agarwood double linked 2-(2-phenylethyl)chromones.

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No.	Name	Source	Ref.
205	(–)-Aquisinenone C	A. sinensis	150
206	(+)-Aquisinenone C	A. sinensis	150
207	(+)-6"-hydroxy-4',4""-dimethoxyaquisinen- one B	A. sinensis	150
208	(+)-Aquisinenone B	A. sinensis	150
209	(–)-6"-hydroxyaquisinenone B	A. sinensis	150
210	(–)-Aquisinenone B	A. sinensis	150
211	Aquisinenone P	A. crassna	151
212	Aquisinenone Q	A. crassna	151
242		A. sinensis	150
213	(+)-Aquisinenone A	A. crassna	151
214	(–)-Aquisinenone A	A. sinensis	150
214		A. crassna	151
215	(–)-4'-methoxyaquisinenone A	A. sinensis	150
216	(–)-Aquisinenone D	A. sinensis	150
210		A. crassna	151
217	(–)-4'-demethoxyaquisinenone D	A. sinensis	150
217		A. crassna	106
218	(+)-4'-demethoxyaquisinenone D	A. sinensis	150
		A. crassna	106
219	3'-hydroxyaquisinenone D	A. crassna	122
220	Aquisinenone R	A. crassna	151
221	(+)-Aquisinenone D	A. crassna	151
222	Crassin H	A. crassna	148
223	(–)-Aquisinenone F	A. sinensis	150
224	(+)-Aquisinenone E	A. sinensis	150
224a	Crassin I	A. sinensis	147
224b	Crassin J	A. sinensis	147
224c	Crassin K	A. sinensis	147

 Table 17: Double linked 2-(2-phenylethyl)chromones with C–O–C and C–C bonds.

# 2.2.3. Sesquiterpenoid-4H-chromones (STCc) and benzylacetone-4H-chromones (BACs)

It is reported that the rare sesquiterpenoid-4H-chromone derivatives (STCs, **225–234**) are reported from the species of agarwood (Figure 20, Table 18). These STCs are composed of a PEC (unit A) and a sesquiterpene moiety (unit B) linked together by an ester bond or an ether bond (Figure 20, Table 18). The *A. crassna* agarwood collected in Laos contains STCs of **225–230**, which are composed by the coupling of a sesquiterpene moiety (unit B) at the C-8 position of the

TPEC (unit A) by an ester bond [152]. The only qinanmer STC **231** is a sesquiterpenoid-4H-chromone reported from the Chinese agarwood "Lv Qi-Nan" of *A. sinensis*.<sup>126</sup> The Cambodian variety of A. crassna agarwood resulted the STCs of **232–234**, which are formed by the sesquiterpene moiety connected to the EPEC through an ether bond [151]. Furthermore, the benzylacetone-4*H*-chromone derivatives **235** and **236** are reported from the agarwood of *G. salicifolia* [137]. These STCs are consisting an unusual 3,4-dihydro-2*H*-pyran ring, which is formed by a C–O–C bond and a C–C bond at C-7 and C-5, respectively (Figure 20).



Figure 20: Sesquiterpenoid-4H-chromones and benzylacetone-4H-chromones of agarwood.

No.	Name	Source	Ref.
225	Aquilacrassnin D	A. crassna	152
226	Aquilacrassnin C	A. crassna	152
227	Aquilacrassnin B	A. crassna	152
228	Aquilacrassnin A	A. crassna	152
229	Aquilacrassnin F	A. crassna	152
230	Aquilacrassnin E	A. crassna	152
231	Qinanmer	A. sinensis	126
232	Xcrassin C	A. crassna	151
233	Xcrassin A	A. crassna	151
234	Xcrassin B	A. crassna	151

Table 18: Sesquiterpenoid-4H-chromones from agarwood

# 2.2.4. Trimers

The trimeric 2-(2-phenethyl)chromone compounds,  $AH_{_{19b}}$  (237),  $AH_{_{20}}$  (**238**),  $AH_{_{18}}$  (**239**), and  $AH_{_{19a}}$  (**240**) are reported from the agarwood "Jinko" from Kalimantan (Figure 21 and Table 19). The trimers 237, 239 and 240 are composed of two

TPEC units (unit A and unit B) connected with a 6,7-dihydroxy-2-(2-phenethyl)chromone moiety (unit C) through a 5C–O–6C bond and a 5C–O–7C bond, respectively. The trimer **238** is composed of two TPEC units (A and B) with a 5,8-dihydroxy-2-(2-phenylethyl)chromone (unit C) through a 5C–O–8C bond and a 6C–O–5C bond, respectively (Figure 21).

No.	Name	Source	Ref.
237	AH <sub>19b</sub>	Kalimantan	153
238	AH <sub>20</sub>	Kalimantan	128
239	(5 <i>5</i> ,6 <i>5</i> ,7 <i>R</i> ,8 <i>5</i> )-2-(2-phenylethyl)-6,7,8-trihydroxy-5,6,7,8-tetrahydro- 5-[2-(2-phenylethyl)chromonyl-6,7-dioxy]chromone [AH <sub>18</sub> ]	Kalimantan	143
240	AH <sub>19a</sub>	Kalimantan	153

Table 19: Tri-2-(2-phenylethyl)chromones of agarwood.





Figure 21: Chemical structures of agarwood tri-2-(2-phenylethyl)chromones.

#### 2.2.5. Phenolics and miscellaneous compounds

The volatile oils of agarwood contains various phenylbutanoids including, anisylacetone (**P5**), zingerone (**P6**), and benzylacetone (**P7**) (Figure 22) [29,48,154,155]. These phenylbutanoids generally are substituted with 3', or 4'-OCH<sub>3'</sub> or 4'-OH, or withour any substitution (Figure 22). In this connection, it is interesting to note that the volatile aromatic compounds are reported from the smoke of agarwood, which

might be the degradation products of chromones and lignins. Further, the phenylpropanoid such as 4'-methoxycinnamic acid (**P1**), 4'-methoxy-phenylpropionic acid (**P4**), anisic acid (**P8**), 3-hydroxy-4-methoxy phenylpropionic acid methyl ester (**P9**), and cinnamaldehyde (**P15**) are also reported from agarwood essential oils [24,48,56,65,155,156]. Additionally, the phenolic compounds including syringin (**P10**), and **P11** – **P14** are also reported from the species of agarwood (Figure 22).



Figure 22: Phenolic compounds of agarwood.

On the other hand, the oxygen-containing triterpenoids,  $3\beta$ -olean-12-ene-3,23-diol (**M1**), 3-oxo-22-hydroxyhopane

(M2) and hederagenin (M3) (Fig. 23) are reported from the agarwood of *A. sinensis*[47,53,61,117,155].



Figure 23: Chemical structures of agarwood triterpenoid compounds.

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# 2.3. Chemical constituents of Taiwan agarwood *Excoecaria* formosana

*Excoecaria formosana* (Syn: *Excoecaria crenulata* var. *formosana* Hayata), is a species of flowering plant in the family Euphorbiaceae. It is a shrub and mainly distributed in Tonkin, Indo-China, the southern part of Taiwan in thickets and forests along the seashores. The resin of *Excoecaria* is used as a substitute for agarwood incense. Chemical investigations on this plant resulted in the isolation of structurally diverse compounds. The halimane-type diterpenoids, formosins A–C (**N1–N3**), and and clerodane-type diterpenoids formosins D–F (**N4–N6**), are isolated from the 95% ethanolic extract of the twigs of *E. formosana* [157]. The whole plant of *E. formosana* led to the isolation of various metabolites; one apocarotenoid, seven benzenoids, cerebrosides, three coumarins, six coumarinolignans, three diterpenes, two flavonoids, six steroids, and eight galloyl glucosides (**N7–N50**, Figure 24, Table 20) [158].



Figure 24: Chemical structures of Excoecaria formosana constituents.

No.	Name	Source	Ref.
N1	Formosin A	E. formosana	157
N2	Formosin B	E. formosana	157
N3	Formosin C	E. formosana	157
N4	Formosin D	E. formosana	157
N5	Formosin E	E. formosana	157
N6	Formosin F	E. formosana	157
N7	$7\alpha$ -hydroperoxysitosterol-3- $O$ - $\beta$ -D-(6- $O$ -palmitoyl)glucopyranoside	E. formosana	158
N8	Excoecoumarin A	E. formosana	158
N9	Excoecoumarin B	E. formosana	158
N10	Excoeterpenol A	E. formosana	158
N11	Deglucosyl lauroside B	E. formosana	158
N12	Gallic acid	E. formosana	158
N13	Methyl gallate	E. formosana	158
N14	4-methoxybenzoic acid	E. formosana	158
N15	3-hydroxy-1-(3,5-dimethoxy-4-hydroxyphenyl)propan-1-one	E. formosana	158
N16	3-hydroxy-1-(4-hydroxy-3-ethoxyphenyl)propan-1-one	E. formosana	158
N17	2,3-dihydroxy-1-(4-hydroxy-3-methoxyphenyl)propan-1-one	E. formosana	158
N18	(2 <i>S</i> ,3 <i>R</i> )-4 <i>E</i> -dehydrochebulic acid trimethyl ester	E. formosana	158
N19	Gynuramide I	E. formosana	158
N20	Gynuramide II	E. formosana	158
N21	Gynuramide III	E. formosana	158
N22	Gynuramide IV	E. formosana	158
N23	Scopoletin	E. formosana	158
N24	Fraxetin	E. formosana	158
N25	6-hydroxy-5,7-dimethoxycoumarin	E. formosana	158
N26	Cleomiscosins A	E. formosana	158
127	Cleomiscosins B	E. formosana	158

N28	Cleomiscosins C	E. formosana	158
N29	Cleomiscosins D	E. formosana	158
N30	Malloapelin A	E. formosana	158
N31	Malloapelin B	E. formosana	158
N32	<i>ent</i> -11 <i>a</i> -hydroxy-3-oxo-13 <i>-epi</i> -manoyloxide	E. formosana	158
N33	Excoecafolin D	E. formosana	158
N34	Agallochin I	E. formosana	158
N35	(+)-catechin	E. formosana	158
N36	Kaempferol-3- <i>O</i> -β-D-glucoside	E. formosana	158
N37	6'-(stigmast-5-en-7-one-3- $O$ - $\beta$ -glucopyransidyl)hexadecanoate	E. formosana	158
N38	(6'- <i>O</i> -palmitoyl) sitosterol-3- <i>O</i> -β-D-glucoside	E. formosana	158
N39	eta-sitosterol	E. formosana	158
N40	Stigmasterol	E. formosana	158
N41	3- <i>O-β</i> -D-glucopyranosyl β-sitosterol	E. formosana	158
N42	3- $O$ - $\beta$ -D-glucopyranosyl stigmasterol	E. formosana	158
N43	lsopropyl O- $\beta$ -(6'-O-galloyl)glucopyranoside	E. formosana	158
N44	4-hydroxy-3-methoxyphenol 1- $O$ - $\beta$ -D-(2',6'-di- $O$ -galloyl)glucoside	E. formosana	158
N45	3-methoxy-4-hydroxyphenyl 1-O- $\beta$ -D-(6'-O-galloyl)glucopyranoside	E. formosana	158
N46	1,2,3,4,6-penta-O-galloyl-β-D-glucose	E. formosana	158
N47	Corilagin	E. formosana	158
N48	1,4,6-tri-O-galloyl-β-D-glucose	E. formosana	158
N49	1,3,6-tri-O-galloyl-β-D-glucose	E. formosana	158
N50	Gallic acid 4- <i>O</i> -β-D-(6'-O-galloyl)-glucose	E. formosana	158

**Table 20:** Chemical constituents of *Excoecaria formosana*.

## 3. PHARMACOLOGICAL ACTIVITIES OF AGARWOOD

The agarwood-isolated compounds/extracts showed various pharmacological activities including, anti-inflammatory, anti-allergic, anti-diabetic, anti-cancer, anti-oxidant, anti-ischemic, anti-microbial, and effects on the central nervous system [3,6,12,25]. The details are described below.

# 3.1. Anti-inflammatory activity of agarwood compounds/ extracts

Inflammation is a vital biological phenomenon that occurs in

response to internal and external injurious stimuli to mitigate foreign triggers, initiate damaged tissue repair and restore the normal body homeostasis [159]. Although the healthy body requires limited inflammation, however, excessive inflammation can cause chronic and degenerative diseases such as diabetes, atherosclerosis, rheumatoid arthritis, cancer and cardiovascular diseases [159]. Nitric oxide (NO) is a proinflammatory mediator that plays a vital role in the process of inflammation [160]. Further, nuclear factor (NF)- $\kappa$ B and tumor necrosis factor (TNF)- $\alpha$  are the key cytokines involved in promoting and triggering the inflammatory process [160].

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Therefore, inhibitors of NO, TNF- $\alpha$ , and NF- $\kappa$ B release may be considered as therapeutic targets for various inflammatory related ailments [160]. Agarwood compounds/extracts are examined for their anti-inflammatory activity through inhibition of NO, TNF- $\alpha$ , and NF- $\kappa$ B release in activated macrophages and neutrophils. Agarwood essential oil has an anti-inflammatory function, significantly reducing the skin thickness, ear weight, oxidative stress, and pro-inflammatory cytokines production in the 12-O-tetradecanoylphorobol-13 acetate (TPA)-induced mouse ear inflammation model [161]. The results of NO inhibitory production are summarized in Table 21.

The sesquiterpenoids, **E17, E26** and **E28** (Figure 7), are reported as inhibitors of NF-kB activation in the activated RAW264.7/Luc-P1 cell line, however, these compounds had not affect the NO release in LPS activated RAW264.7 macrophages (Table 21). Further, the sesquiterpenoids **D4** (Figure 6), and **E28** (Figure 7), at a concentration of 50  $\mu$ M, suppress the superoxide anion generation in fMLP-activated human neutrophils [53]. On the other hand, among the reported

PECs, the compounds 11, 16 and 48 (Figure 14), inhibited the LPS-induced NO production in RAW264.7 macrophages and NF-kB activation in the RAW264.7/Luc-P1 cell line [62]. The authors reported that the presence of a 6-methoxy moiety increased the activity, while the C4' hydroxylation decreased. The PECs 9, 37, 47 and 75 reduced the release of TNF- $\alpha$  in LPSactivated RAW264.7 cells.93 Further, the PECs 6, 26, 52, 60, 66 (Figure 14), and 146 (Figure 16), suppressed the superoxide anion generation in fMLP-activated human neutrophils [53]. The structure-activity relationship (SAR) analysis indicated that the C2'-hydroxy and C4'-methoxy enhanced the activity. Furthermore, the chlorinated PEC 127 (Figure 15), reduced the expression of various inflammatory mediators, such as iNOS, COX2 TNF-α, IL-6, IL-1β, and PGE2 in LPS-activated RAW264.7 macrophage. A mechanistic study revealed that compound 127 selectively suppressed phosphorylation of STAT1/3 and ERK1/2 and activation of NF-kB/MAPK/STAT pathways [162]. A recent study reported that the alcohol extracts of agarwood alleviates the occurrence and development of gastric ulcers via inhibiting oxidation and inflammation [163].

No.ª	NO inhibition (IC <sub>50</sub> , μM)	Ref.	No.ª	NO inhibition (IC <sub>50</sub> , μM)	Ref.	No.ª	NO inhibition (IC <sub>50</sub> , μM)	Ref.
D5	7.2	31	99	5.12	107	192	1.6	139
D8	7.1	31	107	4.5	91	193	5.8	139
D12	3.2	30	111	7.71	107	194	8.1	139
D18	12.8	31	112	3.8	91	195	0.6	139
D24	14.2	30	114	13.09	107	196	0.7	139
D43	2.5	30	117	22.6	107	203	8.0	150
E7	53.8	31	123	22.26	107	204	11.4	150
E9	9.3	31	124	9.01	107	207	10.5	150
E25	12.5	30	126	7.3	91	208	8.8	150
E37	17.3	30	128	4.5	91	210	8.6	150
F33	8.1	32	138	1.6	91	213	11.5143	150
14	6.4	91	140	84	113	214	7.6	150
28	5.95	107	159	7.6	139	215	9.3	150
45	7.59	107	160	7.4	139	216	7.0	150
51	4.6	113	161	2.3	139	217	8.5	150
73	7.94	107	174	37.1	139	218	8.5	150
82	6.59	107	188	4.3	139	223	12.0	150
85	7.94	107	191	1.8	139	D56	5.46	26

D57	14.07	26	D59	45.49	26	B13	52.25	26
B14	62.57	26	F43	66.0	69	F44	76.8	69
F45	62.7	69	F46	18.8	69	F47	72.8	69
G6	89.5	69	G7	68.5	69	G8	74.8	69
G9	84.3	69	135a	3.46	121	135b	12.52	121
135c	> 40	121	162a	35.45	121			

<sup>a</sup>Compound number

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Table 21: Inhibitory effect of agarwood compounds on LPS-induced nitric oxide (NO).

## 3.2. Cytotoxic potential of agarwood compounds/extracts

Agarwood compounds are tested to examine their cytotoxicity in various cancer cell lines such as A549 (human lung), human hepatoma carcinoma cell lines, BEL-7402 and SMMC-7721, Hela (human cervical), K562 (human myeloid leukemia), KB (epidermoid carcinoma), KB-VIN [vincristine (VIN)-resistant KB], MGC-803 (human gastric cancer), OV-90 (human ovarian), breast cancer cell lines, MCF-7 and MDA-MB-231, and SGC-7901 (human gastric). The tested compounds showed weak or moderate cytotoxicity (Table 22).

No.ª	IC <sub>50</sub> (cell line)	Ref.
E11	17.85 μg/mL (K562), 21.82 μg/mL (BEL-7402)	60
F32	33.8 μM (K562)	68
F36	45.1 μΜ (K562)	68
F38	48.6 μM (K562)	68
6	26.2 μM (A549), 19.2 μM (KB-VIN)	84
17	47.0 μΜ (K562), 37.95 μg/mL (SMMC-7721), 35.25 μg/mL (MGC-803), 26.98 μg/mL (OV- 90), 33.8 μΜ (A549), 36.6 μΜ (KB-VIN), 29.0 μΜ (MCF-7)	54,84,8
19	18.1 μM (K562), 20.1 μM (BEL-7402)	54
33	31.59 μg/mL (SMMC-7721), 33.12 μg/mL (MGC-803), 30.77 μg/mL (OV-90)	81
41	13.20 μΜ (K562), 25.91 μΜ (BEL-7402), 23.51 μΜ (SGC-7901), 22.00 μΜ (A549), 30.55 μΜ (HeLa)	116
46	45.38 μM (K562), 35.42 μM (SGC-7901), 33.31 μM (A549)	116
48	22.21 μM (SGC-7901), 8.36 μM (K562), 5.76 μM (BEL-7402)	54, 114
55	30.01 μg/mL (SMMC-7721), 35.25 μg/mL (MGC-803), 26.98 μg/mL (OV-90)	81
56	61.31 μM (K562), 28.53 μM (BEL-7402), 17.63 μM (SGC-7901), 49.42 μM (HeLa)	102
57	37.64 µM (SGC-7901), 27.08 µg/mL (SMMC-7721), 31.17 µg/mL (MGC-803), 33.51 µg/mL (OV-90)	81, 114
58	21.40 μg/mL (SMMC-7721), 36.42 μg/mL (MGC-803), 35.38 μg/mL (OV-90)	81
60	43.65 μg/mL, 14. 96 μM (K562)	106, 10
61	17.8 μΜ (SGC-7901), 13.9 μΜ (K562), 31.9 μΜ (BEL-7402), 25.8 μΜ (A549), 26.1 μΜ (KB), 21.9 μΜ (KB-VIN), 38.1 μΜ (MDA-MB-231), 28.7 μΜ (MCF-7)	54, 84
62	18.82 μg/mL (SMMC-7721), 25.35 μg/mL (MGC-803), 31.60 μg/mL (OV-90)	81
65	20.01 μg/mL (SMMC-7721), 31.34 μg/mL (MGC-803), 36.64 μg/mL (OV-90)	81

70	
78	24.85 μg/mL (SMMC-7721), 28.60 μg/mL (MGC-803), 30.40 μg/mL (OV-90)
81	31.06 μg/mL (SMMC-7721), 28.24 μg/mL (MGC-803), 22.54 μg/mL (OV-90)
84	11.83 μM (K562), 25.02 μM (BEL-7402), 29.29 μM (SGC-7901), 44.11 μM (HeLa)
110	35.11 μM (BEL-7402), 32.95 μM (SGC-7901)
134	14.6 mg/mL (SGC-7901)
136	46.1 μM (SGC-7901), 43.8 μM (A549)
152	2.87 μΜ (K562), 4.75 μΜ (BEL-7402), 9.91 μΜ (SGC-7901), 22.43 μΜ (A549), 13.86 μΜ (HeLa)
153	40.81 μM (K562), 44.18 μM (BEL-7402)
175	73.5 μM (K562)
182	70.9 μM (K562)
199	44.68 μM (BEL-7402)
200	42.10 μM (BEL-7402)
213	34.20 μM (SGC-7901), 37.99 μM (K562), 36.26 μM (HeLa)
214	11.59 μM (SGC-7901), 22.97 (A549), 10.93 μM (K562), 12.88 μM (HeLa)

33.9 μM (K562), 29.9 μM (BEL-7402), 26.7 μM (HeLa), 46.3 μM (A549)

39.95 µM (SGC-7901), 28.67 µM (K562), 29.34 µM (HeLa)

24.8 μM (BEL-7402), 30.9 μM (SGC-7901), 17.6 μM (HeLa), 32.0 μM (A549)

31.50 μM (SGC-7901), 49.0 μM (A549), 22.12 μM (K562), 30.75 μM (HeLa)

<sup>a</sup>Compound number

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Table 22: Cytotoxicity potential of agarwood compounds in various cancer cell lines.

## 3.3. Neuronal activity of agarwood compounds/extracts

25.7 μM (BEL-7402), 30.6 μM (HeLa)

It is known that agarwood traditionally used as a sedative and analgesic agent [3]. The pharmacological studies reported that agarwood extracts as well as pure compounds showed neuroprotective activity [3,6,12]. For example, the benzene extract of A. malaccensis agarwood reduced spontaneous motility, prolonged hexobarbiturate-induced sleeping time, and decreased rectal temperature, while the petroleum ether, chloroform, or water extracts did showed the similar effect [164]. A bio-quided isolation of a benzene extract yielded the jinkoh-eremol (E3, Figure 7) and agarospirol (B1, Figure 4) are the main active constituents [165,166]. The agarwood essential oil sedated mice through vapor inhalation, and identified the main volatile compounds are benzylacetone,  $\alpha$ -gurjunene, and (+)-calarene [167]. The 70% EtOH extract of Vietnamese agarwood induced the expression of brain-derived neurotrophic factor (BDNF) mRNA in rat cultured neuronal cells, and identified the sesquiterpene B5

(Figure 4), is responsible active compound to the observed biological potential [36]. The PEC compound 73 (Figure 14), showed neuroprotective activity in P12 pheochromocytoma, and human U251 glioma cells against glutamate-, and corticosterone-induced neurotoxicity [115]. The alcohol extract of agarwood produced by whole-tree agarwoodinducing technique, and the volatile oil combined with pentobarbital sodium showed hypnotic effect. These tested agents prolonged the sleeping time, and increased the rate of falling asleep in mice [168]. In addition, it is also found that the agarwood essential oil showed sedative-hypnotic effects through the GABAergic system [169]. Agarwood essential oil ameliorates restrain stress-induced anxiety and depression by inhibiting HPA axis hyperactivity [170]. The diterpenoids of agarwood showed antidepressant activity through the synaptic reuptake of serotonin and norepinephrine [171]. A recent study reported that the low molecular weight aromatic compounds (LACs) obtained from the headspace-solid phase

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microextraction (HS-SPME) of *Kyara* grade (highest-grade agarwood in Japan), showed strongest sedative activity in mice [172]. Agarwood smoke from Kynam agarwood, showed anti-anxious and anti-depressant effects associated with the increase of serotonin levels in mice [173]. Furthermore,

agarwood compounds are reported as as promising therapeutic agents to combat Alzheimer's disease through inhibition of acetylcholinesterase (AChE) activity (Table 23). The AChE inhibitory potential of agarwood compounds are presented in Table 23.

No.ª	Inhibition rate(%)	Ref.	No.ª	Inhibition rate (%)	Ref.	No.ª	Inhibition rate (%)	Ref.
D22	21.2	57	2	24.1	80	81	19.6	114
E15	33.3	41	3	14.3	80	82	21.6	41
E16	274.8 μM (IC <sub>50</sub> )	58	6	19.3	88	83	41.47	120
E18	32.7	57	8	15.8	89	84	33.6	88
E26	491.4 μM (IC <sub>50</sub> )	58	9	17.4	89	87	41.27	120
E28	158.3 μM (IC <sub>50</sub> )	58	14	38.0	111	88	32.11	120
E29	42.9	40	20	11.4	41	100	17.5	127
E33	15.2	57	21	20.3	83	105	10.61	37
F17	19.5	71	27	16.3	80	125	19.1	127
F19	19.4	71	28	23.5	88	135	21.10	37
F21	19.1	67	32	10.0	80	139	31.5	88
F22	63.1	59	35	17.0	80	142	15.8	118
F23	15.0	67	33	14.9	80	143	35.9	118
F26	24.1	67	37	26.9	80	146	47.9	127
F30	31.0	71	38	25.4	83	148	155.6μM (IC <sub>50</sub> )	135
F35	54.2	41	39	24.0	41	149	441.6μM (IC <sub>50</sub> )	135
F40	35.3	41	40	10.8	100	151	47.4	127
F41	46.2	41	42	10.1	88	163	16.82	142
<b>B6</b>	16.35	37	44	12.2	114	167	44.01	145
<b>C</b> 1	49.9	39	47	15.0	80	171	10.85	145
A1	44.5	71	60	22.0	106	172	24.57	145
A2	20.8	71	62	35.0	109	173	16.80	142
1	18.6	80	65	10.0	83	185	15.66	142
E41	48.33	50						

<sup>a</sup>Compound number

Table 23: Acetylcholinesterase inhibitory activity of agarwood compounds (at 50 µg/mL).

# 3.4. Anti-diabetic activity of agarwood compounds/ extracts

Diabetes mellitus (DM), known as diabetes is a serious, chronic, and complex metabolic disorder [174]. The DM complications affect people both in the developing and developed countries. There are several classes of therapeutic antidiabetic drugs such as sulfonylureas, biguanides,  $\alpha$ -glucosidase inhibitors, thiazolidinediones, and non-

sulfonylureas secretagogues [174]. The agarwood compounds are reported as  $\alpha$ -glucosidase inhibitors. For example, A. filaria sesquiterpenoid, guaianolide (**F37**) reported as an inhibitor of  $\alpha$ -glucosidase with an IC<sub>50</sub> value of 253.2  $\mu$ M [68]. Further, the prezizaane sesquiterpenoids, **H1**, **H4**, and **H11** (Figure 10), and zizaane sesquiterpenoids, **I1** and **I3** (Figure 11), are reported to possess the inhibitory effect against  $\alpha$ -glucosidase [76]. On the other hand, the PECs compounds **11** and **12** (Figure 14), shown to promote the secretion of adiponectin as PPARy agonists

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during adipogenesis in human bone marrow mesenchymal stem cells [82]. The *A. sinensis* PEC compounds **47, 48** and **65** (Figure 14), reported as inhibitors of  $\alpha$ -glucosidase with IC<sub>50</sub> values of 90, 50 and 150  $\mu$ M, respectively [114].

# 3.5. Antibacterial activities of agarwood compounds/ extracts

The sesquiterpenoids and 2-(2-phenylethyl) chromones (PECs) of *A. crassna* and *A. sinensis* are examined for their antibacterial activity against *Staphylococcus* aureus and *Ralstonia* 

solanacearum using disk agar diffusion method (Table 24). The PECs of *A. sinensis* agarwood showed antibacterial activity against *S. aureus*, and methicillin-resistant *S. aureus* (MRSA) (Table 24) [99]. The sesquiterpene  $\beta$ -caryophyllene (G5, Figure 9), showed superior antibacterial activity against Gram-positive human pathogenic bacteria than that of Gram-negative bacteria [73]. The extracts of *A. crassna* agarwood, aqueous, SFE, and SFE with ethanol as the co-solvent, are showed antimicrobial activities against *S. aureus* and *Candida albicans*, but are not against *Escherichia coli* [175].

No.ª	S. aureus	R. solanacearum	Ref.
D4	20.02	11.02	52
D5	9.12	8.98	52
D8	12.90	18.20	52
D9	14.20	10.15	52
D10	8.10	Not active	52
D31	12.35	16.90	40
6	Not active	6.80	89
54	9.10	Not active	88
56	10.01	Not active	88
145	14.95	12.09	88
146	12.75	15.40	88
P4	11.20	7.81	155

<sup>a</sup>Compound number

Table 24: Antibacterial activity (Inhibition zone in mm) of agarwood compounds

# 3.6. Effect of agarwood compounds/extracts on cardiovascular System

It is reported that 50% ethanolic extract of Bawei Chenxiang powder enhanced the hypoxia tolerance of cardiomyocytes [176]. The Tibetan Bawei Chenxiang powder showed a protective effect on the ratmodel of myocardial ischemia [177]. The agarwood alcohol extract ameliorates isoproterenol-induced myocardial ischemia by inhibiting oxidation and apoptosis [178]. The agarwood of *A. crassna* showed noticeable cardioprotective activities. For example, *A. crassna* extract reduced simulated ischemia induced cell death in cardiac myoblast cell line, H9c2 [179], as well as isolated adult rat ventricular myocytes [180]. Additionally, the ethyl acetate extract of *A. crassna* protect the heart from myocardial

ischemia/reperfusion injury through attenuation of p38 MAPK phosphorylation [181]. Further, it is also reported the cytoprotective effect of *A. crassna* extract on actin cytoskeleton organization, in cardiac cell subjected to simulated ischemia [182]. Phosphodiesterases (PDEs) are enzymes that regulate cellular signaling by hydrolysis of intracellular second messengers, cyclic adenosine monophosphate (cAMP), and cyclic guanosine monophosphate (cGMP) [183]. In cardiovascular tissues, PDE 3A is one of the dominant cAMP-hydrolyzing isozymes, and PDE 3 inhibitors may be used in congestive heart failure [183]. On the other hand, PDE 5 is the major GMP hydrolyzing enzyme in human corpus carvernosal tissue, and PDE 5 inhibitors such as sildenafil have been used to treat erectile dysfunction [183]. A recent study reported

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that the new FPEC (**85a**, Fig. 14) and DIPECs **198a–198d** (Figure 18), have considerable activity against PDE (Table 25).<sup>79</sup> Additionally, the PECs compounds **19, 20, 47, 48, 56, 60** and

**81** (Figure 14), are also reported to have PDE 3A inhibitory activity [104].

No.ª	PDE	IC <sub>50</sub>	Ref.
19	PDE 3A	89.3 µM	79
	PDE 5A1	19.4 µM	79
48	PDE 3A	4.83 µM	104
85a	PDE 3A	44.2 µM	79
	PDE 5A1	20.7 µM	79
198a	PDE 3A	> 100 µM	79
	PDE 5A1	4.2 μM	79
198b	PDE 3A	> 100 µM	79
	PDE 5A1	7.9 μΜ	79
198c	PDE 3A	42.6 µM	79
	PDE 5A1	15.1 μM	79
198d	PDE 3A	> 100 µM	79
	PDE 5A1	4.3 µM	79

<sup>a</sup>Compound number

Table 25: Effect of agarwood compounds against PDE activity.

#### 3.7. Other activities

The sesquiterpenoid **D8** (Figure 6) reported as inhibitor of innate and adaptive immunity through suppressing the STAT signaling pathway [184]. The PECs 18, 65, 75 (Figure 14), and 158 (Fig. 17), showed ABTS<sup>++</sup> radical scavenging activity, with IC<sub>50</sub> values of 34.7, 16.5, 12.1 and 12.3  $\mu$ M, respectively [92]. The A. sinensis PEC compound 136 (Figure 16), suppressed the survival, activation, proliferation, and differentiation of B cells through reduced B-cell activating factor from the tumor necrosis factor family (BAFF) signaling [185]. The PEC compounds 16, 44 and 82 (Fig. 14), showed the tyrosinase inhibitory activity [75,103]. The ethanolic extract of agarwood produced by the whole-tree agarwood-inducing technique, improved the intestinal peristalsis, enhanced gastric emptying, and inhibited gastric ulcer [175]. Additionally, agarwood ethanol extract showed protective effect of intestinal injury induced by fluorouracil (5-FU) through reduced inflammation and, enhanced antioxidant enzymes and Nrf2 signalling [186]. The alcoholic extract of Agar-Wit agarwood alleviate the inflammation and asthma in the asthma mouse model

induced by intraperitoneal injection of ovalbumin+Al(OH)<sub>3</sub> [187]. The agar wood decotions/infusions traditionally used for alleviating abdominal discomfort, however the gastrointestinal effect on a specific disease is not completely explored yet.

# 3.8. Biological activities of *Excoecaria formosana* compounds

Formosins F (**N6**, Figure 24) showed moderate anti-microbial activity against two strains of *Helicobacter pylori* (Hp-SS1 and ATCC 43504) with MIC values of 50 and 50 µg/mL, respectively [157]. Compounds **N44**, and **N46–N48** (Figure 24), at a 100 µM concentration showed a 2.97-, 3.17-, 2.73-, 2.63-, 6.57, and 2.62-fold increase in glycine N-methyltransferase (GNMT)-promoter activity, respectively [158].

### 3.9. Clinical Application of agarwood

The clinical studies indicating that agarwood has therapeutic effect in various diseases, including cardio-cerebrovascular system, urinary system, and respiratory system. The agarwood

product Bawei Chenxiang powder (agarwood, nutmeg, jujube, travertine, frankincense, radix aucklandiae, chebula, kapok), showed therapeutic effect in patients with bronchial asthma [188]. Additionally, the Bawei Chenxiang powder showed superior protective effect in the patients with angina pectoris (a coronary heart disease), after continuous administration for four weeks, as compared with the conventional medicine treated patients [189]. In an another study 30 constipation patients are treated with Chenxiang Tongbian powder. The results showed that the total effective rate of Chenxiang Tongbian powder is higher (13.33%) than that of patients treated with polyethylene glycol electrolyte orally, and the symptoms of the patients were significantly relieved after two weeks of treatment [190].

## 4. EXTRACTION AND ANALYSES OF AGARWOOD

#### 4.1. Extraction of agarwood

In general, the agarwood extraction method depends on the purpose of the extract [191]. The agarwood essential oils are obtained through hydrodistillation, or steam distillation [191]. The chemical constituents of agarwood usually obtained from the solvent extraction, such as acetone, methanol, ethanol and water or supercritical fluid extraction [191]. Various extraction process such as maceration, soxhlet, supercritical fluid, ultrasonic-assisted, microwave-assisted, and highpressure processing extractions are used to get the desired extract/compounds [191]. Each solvent and/or extraction process produces different extracts in terms of quantity and quality of the constituents [191]. Although water is a cheap solvent and relatively safe, however, aqueous extracts resulted the impurities that makes difficult to isolate the desired compound. Therefore, after the aqueous extraction process, the crude extract was fractionated with hydroalcohol into the desired compounds [191]. This technique is widely applied, especially in the whole process of extraction of the agarwood [191]. Methanol also suitable as an extraction solvent since aqueous methanol was more effective in extracting total sesquiterpenes, 2-(2-phenylethyl)-4H-chromen-4-one derivatives (PECs), and aromatic compounds as compared with water [191]. Alternatively, ethanol is also a suitable solvent for agarwood extraction. It is a non-toxic for human consumption, and widely used for natural products extraction [191]. Most secondary metabolities are dissolved in ethanol except protein, phlegm, pectin, starch and polysaccharide [191].

It is difficult to distinguish agarwood quality by observing its morphological characteristics, and the handicrafts of incense are more complicate to identify. Recently, the chemical constituents of agarwood has gained increasing attention due to the agarwood quality is correlate its resin yield and metabolites [72,192]. At present, various methods are established to control the quality of agarwood, including the coloration of chemical reagents, the content of alcohol extracts, the content of agarwood agarotetrol (98, Figure 15), the content of chromone, the HPLC fingerprint of alcohol extract, etc. Using these methods, it is conceivable to clarify the agarwood obtained either from wild or artificial induction methods [193,194]. The Chinese Pharmacopoeia (2020 edition) indicates that the content of the ethanol extract of agarwood resin needs to be more than 10% (w/w, dry weight), and the content of the marker compound, agarotetrol (98, Figure 15) needs to be more than 0.1% [10].

### 4.2.1. Analyses of agarwood sesquiterpenoids

Several analytical techniques are applied to analyze the agar wood chemical compounds of essential oils, such as electronic nose (Enose), gas chromatography (GC), GC/mass spectrometry (GC/MS), solid phase micro extraction (SPME), GC-flame ionization detector (GC-FID), GC-olfactometry (GC-O), and comprehensive two dimensional GC [191]. Among these, GC/MS, followed by SPME techniques are preferential, which showed promising result in analysing the chemical compounds of agarwood oil [191]. The sesquiterpenoids from the agarwood of A. agallocha and A. malaccensis are identified using the combination of GLC and GC/MS [191]. It is reported that the abundances (percentage of relative peak area measured by GC-MS) of the same compound in high quality oil is more than that of low quality agarwood oil [196]. Further, Ishihara et al., (1993), classified the quality of agarwood oil based on peak area percentage (or abundances) of α-guaiene (F8, Figure 8) with the peak area < 0.05% classified as a high quality oil, however the wood oil which is not containing α-guaiene (F8) classified as low guality [65]. Moreover, lower quantity of benzaldehyde (P9, Figure 22) and anisaldehyde (P10, Figure 22) are present in high grade agarwood oil (i.e. Kanankoh), as compared with the low grade (i.e. Jinkoh), agarwood oil [65]. Additionally, the same authors also reported that the quantity of agarwood resin and content of oxygenated sesquiterpenes are comparatively higher in high grade agarwood oil [42]. Table 26 summarizes the component based characteristic of agarwood oil. It is reported that the GC-MS analysis of aromadendrene (F42, Figure 8), showed a

### 4.2. Analyses of agarwood

positive linier relationship with the agarwood resin yield and quality, and therefore suggests as a marker compound for agarwood grading [72]. The eremophilane-type sesquiterpene, valencene (**E39**, Figure 7) from *A. malaccensis* is reported as a marker compound in the grading of agarwood oil [64].

Component	High Quality	Low Quality	Ref.
α-guaiene ( <b>F8</b> )	<0.05%	Not available	19,65
Resin content	high	low	197
Benzaldehyde ( <b>P9</b> ), anisaldehyde ( <b>P10</b> )	Less amount	More amount	65,198
10-epi-γ-eudesmol ( <b>D21</b> ), β-agarofuran ( <b>D44</b> ), α-agarofuran ( <b>D49</b> )	Presence	Not mentioned	154
β-agarofuran ( <b>D44</b> )	Marker compound	Not mentioned	199

Table 26: Component based characteristic of agarwood oil.

# 4.2.2. Analyses of agarwood 2-(2-phenylethyl)chromens (PECs)

On the other hand, the PECs compounds are the major fragrance constituents of agarwood, which contributors to the sweet, fruity and long lasting scent of agarwood when it is burn [3,6,12]. The content of agarwood PECs are used to evaluate the grading of agarwood products [86]. Various types of agarwood specific PECs are identified as potential marker for its authentication [25]. These PEC compounds are obtained through solvent extraction methods, but are not extractable using hydrodistillation [200,201]. Structural studies revealed that most of the reported agarwood PEC compounds has the same basic skeleton (MW: 250) and similar substituents, i.e., hydroxy or methoxy or both groups [202]. The determination of PECs in agarwood using GC-MS are relatively limited, as compared with the sesquiterpene constituents. It is reported that agarwood "Kanankoh" oil as a

high-grade one, while the "Jinkoh" agarwood is the low quality [42,65,197]. The "Kanankoh" oil contains 66.47 % of PECs and 2-(2-4-methoxy-phenylethyl) chromone, which is higher than the "jinkoh" (1.5%) [196]. The contents of PECs, agarotetrol (98, Figure 15) and isoagarotetrol (103, Figure 15), have positive correlation with the quality of commercial agarwood [203]. An integrated strategy using SHS-GC-MS and UPLC-Q/Tof-MS is used to discriminate the high grade wild Chi-Nan agarwood from A. sinensis, and ordinary agarwood. The results revealed that average contents of 2-(2-phenylethyl) chromones and sesquiterpenes in Chi-Nan agarwood is higher as compared with ordinary agarwood [204]. A recent study reported that the pharmacokinetic results of major PECs in rat plasma using UHPLC with tandem mass spectrometry after oral administration of agarwood ethanol extract [205]. It is interesting to note that HPLC is a superior analytical technique to analyze the agarwood PECs. The details are presented in Table 27.

Species	Sample	Extraction method	Column / Temerature	IV <sup>a</sup> /DW <sup>b</sup>	Mobile phase/ runtime (min)	FLº (ml/min) / Compounds	Ref.
Agarwood spp.	AW	EtOAc fract. of 95%EtOH	Gemini RP-C18 (250 × 10, 5 μm)	5/254	CH <sub>3</sub> OH–H <sub>2</sub> O (70:30) / 25	PECs	162
Agarwood spp.	AW	alcohol	Diamonsil C18 (250 × 4.6, 5 μm)/ 32°C	10/252	0.7 /	PECs	206
Agarwood spp	Wild, cultivar		Phenomenex Luna C18 150×4.6,5mm/31°C	10/252	CH <sub>3</sub> CN - 0.1% HCOOH / 60	0.7/ PECs	207
Agarwood spp	AW	95%ethanol	Diamonsil C18 (4.6 ×250, 5 μm) / 30°C	5 / 252	CH <sub>3</sub> CN, 0.1% HCOOH	0.7 /	208

A. crassna	Trunk	Et <sub>2</sub> O	Dionex-Acclaim 120 C18 (250× 4.6, 5 μm)/ 26°C	20/254	CH <sub>3</sub> CN, CH <sub>3</sub> OOH (99.5:0.5)/ 95	0.4 / PECs	209
A. sinensis	AW	Ultrasonic ext. with Et <sub>2</sub> O	Dionex-Acclaim 120 C18(250 ×4.6,5 µm)/ 26°C	20/254	CH <sub>3</sub> CN, HCOOH (99.5:0.5, / 95	0.4 mL/min/ PECs	210
A. sinensis	AW	MeOH ext	HPLC, QAMS, and UPLC-	PECs	211		
Agarwood spp	HPLC- L-7 mm × 250	Agarotetrol	212				

AW- agarwood; aIV- Injection volume (in µL); DW- Detection wavelength (nm); FL- flow rate; PECs- 2-(2-phenylethyl)chromones

Table 27: HPLC analyses of agarwood samples.

### 4.2.3. Identification of agarwood adulteration

Due to its high demand and high price, agarwood commercial products are tainted with adulteration and substitution products in order to meet the market demand. Agarwood adulteration happened in different forms, such as painting and covering with oil or making the wood heavier, etc. In general, the impregnation of agarwood with abietic acid or wax to create a resemblance to high-grade agarwood [213]. Powder is the most susceptible agarwood item for adulteration, where it is mixed with healthy (uninfected), wood. Two types of fake agarwood have been described; (i) low quality agarwood painted with small layer of shavings mixed with wax and other material; and (ii) "Black Magic Wood" which refers to low quality agarwood impregnated with agarwood oil and alcohol [214]. Iron shavings and carbon powder from spent batteries are also used to increase the weight and create resemblance to high grade agarwood [51]. In Taiwan market, inferior quality of agarwood has been increasingly mis-classified and substituted as the top-grade agarwood. On the other hand, agarwood oil is adulterated either with 'lodh' oil, kerosene, other coloured oils, a mixture of other chemicals that gives the aroma of agarwood [51]. In this connection, various synthetic agarwood compounds are developed [118,212]. However, these are used to produce poor-quality fragrances as no synthetic substitutes are available for high-grade fragrances due to the complexity of natural agarwood composition [51].

Various PECs are specific in agarwood. The DART-TOFMS characteristic fragmentation behavior of PECs is applied for their accurate identification in agarwood [215]. The presence of the diagnostic TPEC ions at m/z 349.129 or at m/z 319.118, are characteristic of the PECs [215]. Interestingly, cultivated agarwood and wild agarwood samples showed differences

in PECs, which helped to distinguish the wild agarwood from cultivated one [216]. The characteristic fragmentation behaviors of FPECs, and cleavage of the CH,-CH, bond between the chromone moiety and phenyl moiety are used to calculate the number of methoxy or hydroxy groups, which enabled the identification of FPECs [202]. Further, the characteristic fragmentation behaviors of DPECs, EPECs and TPECs are analyzed using LC-MS without databases or reference standards [209]. A recednt study reported an integrated method of FT-NIR, GC-MS and UHPLC-Q-Exactive Orbitrap/MS, to identify the chemical variation between wild and cultivated agarwood. This novel method identified eight key marker compounds, including flidersia type (FPECs)-sesquiterpenes and TPECs, which are putatively distinguish between wild and cultivated agarwood [217]. Tian et al., developed an UHPLC-TOF-MS method to compare the chemical composition and the bioactivities of wild and artificial agarwood [218]. The Liquid extraction surface analysis mass spectrometry (LESA-MS) is applied in the direct qualitative analysis of agarwood from different sources [219]. During this study, a characteristic 2-(2-phenylethyl)chromone compound (m/z 319.1) is treated as a marker to identify agarwood and its counterfeits [219]. The headspace GC-MS is used to to distinguish the sesquiterpenes compounds between the high-quality agarwood Kynam, and cultivated grafting Kynam [220].

## **CONCLUSIONS / FUTURE PROSPECTS**

The major compounds of agarwood apecies are sesquiterpenoids, and 2-(2-phenylethyl)chromone. These are diverse and complex, and are influenced by agarwood plant species, formation process, collection time, extraction process, and analytical approach etc. Further studies are required to understand the agarwood resin composition, and to identify

specific marker compounds of different agarwood plant species through metabolomics approach. The agarwood formation mechanism is not completely revealed; therefore, studies are required to understand the essential compounds biosynthesis mechanism of agarwood resin. Although, various pharmalogical activities are reported for the pure compounds/extracts, however, the specific active substances and pharmacological action mechanisms are not been confirmed. Therefore, systematic in vitro and in vivo studies are required for the identification of effective components of agarwood and explore their action mechanism. The active compounds can be used as a template to develop novel therapeutic agents. The relationship between the active compounds and their pharmacokinetics need to be identified to develop their clinical use. Further systematic analytical methods are required to chemically distinguish the different kinds of agarwood. Studies are required to establish an accurate and rapid identification of fake and shoddy products in commercial products.

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